

The role of vitamin D in protecting type 1 diabetes mellitus

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Summary

The relationship between autoimmune diabetes or type 1 diabetes mellitus and vitamin D has been reported in the literature. Many factors, environmental and genetic, have been known, as risk factors, to cause both type 1 diabetes and vitamin D deficiency. Vitamin D treatment has improved or prevented type 1 diabetes mellitus in animals and humans. Vitamin D also has been known to protect from autoimmune diseases in animal models. Therefore, it would be interesting to review the role of vitamin D in type 1 diabetes mellitus. Copyright © 2005 John Wiley & Sons, Ltd.

Keywords vitamin D; diabetes mellitus; 1,25-dihydroxyvitamin D₃

Introduction

Vitamin D has been known as a regulator of bone and mineral metabolism by regulation of calcium absorption in the gut and reabsorption by the kidney. In addition, 1,25-dihydroxyvitamin D₃ [1,25(OH)₂D₃] is also recognized in the regulating of the immune system. Vitamin D receptor (VDR) is presented in peripheral blood monocytes and activated T cells [1,2]. In the animal models, 1,25(OH)₂D₃ protects against autoimmune diseases, such as experimental autoimmune encephalomyelitis (EAE) [3] and collagen-induced arthritis [4]. Recently, Zella and Deluca [5] suggested the effectiveness of vitamin D in diabetes. Therefore, it would be interesting to further review the role of vitamin D in protecting autoimmune diabetes, also known as type 1 diabetes mellitus.

Risk factors for type 1 diabetes mellitus

Many risk factors, environmental and genetic, have been known to contribute in the development of both vitamin D deficiency and type 1 diabetes (Table 1).

Environmental factors

Seasonal and geographical factors have been known, as risk factors, to cause both type 1 diabetes and vitamin D deficiency. There was a suggestion of geographic and seasonal variations for the prevalence of type 1 diabetes. Keen and Ekoe [6] reported that type 1 diabetes is more prevalent in higher latitudes of the tropics and subtropics. In addition, there is a seasonal variation in type 1 diabetes with the largest proportion of cases diagnosed during fall and winter and the lowest during the summer. In 1988, the Diabetes Epidemiology Research International Group [7] studied geographical patterns on risk of type 1 diabetes. This report was compiled from age- and sex-specific cases

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of type 1 diabetes from 1978 to 1980, representing approximately 50 million children younger than 15 years of age from 15 countries of 4 continents. They found that the risk for type 1 diabetes is determined by factor(s) correlated to the average yearly temperature of environment that was strongly associated with latitudes increasing in distance from the equator. Tuomilehto *et al.* [8] analyzed the age/calendar time/birth cohort effects on the increasing trend of type 1 diabetes between 1965 and 1984. They noted that the increase was mainly related to calendar time and that all age groups were similarly affected. Padaiga *et al.* [9] examined seasonal patterns of incidence of type 1 diabetes in children aged 0 to 14 years in the countries around the Baltic Sea (Finland, Sweden, Estonia, Latvia, and Lithuania) during 1983 to 1992 (1987–1992 for Finland). They found that the pattern among younger children (0–9 or 5–9 years) had one cycle with a decreased incidence of type 1 diabetes in May and June. Ishii *et al.* [10] reported a seasonal variation of glycemic control in their diabetic patients. They noted that mean HbA_{1c} levels were elevated by average 0.5% in winter compared to the period between spring and fall. Muntoni *et al.* [11] compared the number of people with type 1 diabetes born in each month of 1974 to 1995 in Finland, where the incidence of type 1 diabetes in children aged <14 years is the highest in the world [7] and there is 2 h of sun every day in December, and Sardinia (an island with very high sunlight exposure). They reported that type 1 diabetes developed at an age younger than 15 years in Finland compared to such children in Sardinia.

Similar seasonal and geographic variations have also been suggested for the prevalence of serum 25-hydroxyvitamin D₃ (25OHD₃) levels. McKenna [12] studied vitamin D status between countries in young adults and in the elderly. He found vitamin D status varies with the season in young adults and in the elderly, and is lower during winter in Europe than in both North America and Scandinavia. Webb *et al.* [13] reported the influence of season and latitude on the cutaneous synthesis of vitamin D₃. In Boston, USA (42.2°N), from November through February, human skin produced no previtamin D₃ when exposed to sunlight on cloudless days. In Edmonton, Canada (52°N), this ineffective period extends from October through March. Further south (34°N and 18°N), sunlight effectively photoconverted dehydrocholesterol to previtamin D₃ in the middle of winter. Fairney *et al.* [14] measured circulating 25OHD₃ levels of Caucasians in the Antarctic. Following a period of 3 months' complete darkness, there was a significant fall in the vitamin levels. Sunlight exposure and vitamin D status are highest in

the summer and lowest during the fall and winter in the northern latitudes [15]. Seasonal variations of 25OHD levels were also reported in Ushuaia (Argentina) (latitude 55°S), the southernmost city of the world [16]. Recently, Barger-Lux and Heaney [17] confirmed and quantified the relatively large seasonal fluctuations in circulating 25OHD₃ levels in association with summer sun exposure among outdoor workers. Median serum 25OHD₃ levels decreased from 122 nmol/L in late summer to 74 nmol/L in late winter.

Recently, there were two articles that investigated dietary exposures in infancy and the risk of developing islet autoantibodies. Norris *et al.* [18] found that the risk of developing islet autoantibodies was significantly higher among children initially exposed to cereal between ages 0 and 3 months or at 7 months than among infants exposed to cereals between ages 4 and 6 months. In the second study, Ziegler *et al.* [19] also reported that introduction of gluten-containing foods before age 3 months was significantly associated with increased risk of developing islet autoantibodies. However, these findings also might suggest a role of vitamin D in their studies. Pileggi *et al.* [20] reported the role of vitamin D in the prevention of rickets in rats on cereal diets. Review of the evidence of the Irish Nutrition Survey [21], concerning a marked rise in the incidence of rickets in Dublin in 1942, concluded that a rise in the extraction rate of the national flour, more phytate and total phosphorus, from 70 to 100% was principally responsible. This rise and, subsequently, decrease in incidence in the extraction rate of flour was reduced. Ford *et al.* [22] suggested a strong and possibly causal relationship between high-extraction cereal and rickets and osteomalacia. Corazza *et al.* [23] reported a change in bone mineral density (BMD), serum calcium, and serum 25-vitamin D₃ levels in gluten-sensitive patients. Gluten-free diet was able to normalize BMD and abnormal blood tests. In addition, the presence of islet cell antibodies has been reported in malnutrition patients [24,25], which is related to rickets. Two of the antigens for islet cell antibodies (ICAs) are glutamic acid decarboxylase (GAD) and a tyrosine phosphatase-like protein termed IA-2. Vitamin D analogs are able to inhibit the spontaneous T-cell response to the intracytoplasmic region of the tyrosine phosphatase-like protein (termed IA-2), an autoantigen associated with type 1 diabetes in both humans and the nonobese diabetic (NOD) mice [26].

The relationship between vitamin D and type 1 diabetes has been known in the past. Baumgartl *et al.* [27] reported that serum 25OHD₃ levels measured at matched time points throughout the year are lower in patients newly diagnosed with type 1 diabetes than in healthy controls. In 1999, the EURODIAB Substudy 2 Study Group [28] studied the correlation between vitamin D supplements during the first year of life with the development of type 1 diabetes. They reported that vitamin D supplement during the first year of life is associated with a decreased risk of type 1 diabetes. In another study, Stene *et al.* [29] investigated whether cod liver oil or vitamin D

Table 1. Similar risk factors for contributing in both vitamin D deficiency and type 1 diabetes

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- Environmental:
 - Season: Fall & winter
 - Geographic: Latitudes distance from the equator
 - Foods: Cereal and gluten-containing foods
 - Genetic: Certain allelic variations in the vitamin D receptor may be of genetic risk for type 1 diabetes.
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supplements taken either by the mother during pregnancy or by the child in the first year of life is associated with lower risk of type 1 diabetes in children. They found a lower risk of diabetes in children when mothers took cod liver oil during pregnancy. It was noted that newborn children of mothers who had taken cod liver oil during pregnancy had higher concentrations of 25OHD₃ in the cord blood than children of mothers who had taken other vitamin D supplements during pregnancy [29]. However, there was no significant protection from type 1 diabetes risks when infants were fed either cod liver oil or vitamin D supplements. They suggested that exposure *in utero* could be relevant for the development of type 1 diabetes. In addition, Hypponen *et al.* [30] assessed the risk of type 1 diabetes and vitamin intake during infancy of 10 821 children in Oulu and Lapland of northern Finland. They reported that dietary vitamin supplementation is also associated with reduced risk of type 1 diabetes.

Genetic factors

Certain allelic variations in the VDR may also be of genetic risk for type 1 diabetes. Several reports now indicate that type 1 diabetes may be included among the list of diseases genetically determined, at least in part, by VDR gene polymorphisms. 1,25(OH)₂D₃ has been shown to inhibit β -cell growth by upregulation of vitamin D receptors, suggesting an alternative mechanism whereby VDR variants might alter insulin responses through early β -cell differentiation [31]. Ortlepp *et al.* [32] tested the influence of the VDR polymorphism on fasting glucose in healthy young men. They found that the VDR genotype is associated with altered fasting glucose levels in young men with low physical activity. McDermott *et al.* [33] found evidence of an association of one particular vitamin D receptor allele with type 1 diabetes susceptibility in Indian Asians. These allelic variations in VDR gene that influence genetic susceptibility to type 1 diabetes have also been reported in other ethnic groups: Brazilian [34], Taiwanese [35], German [36], Japanese [37], Bangladeshi Asians [38], Romanian [39], and Dalmatian population of South Croatia [40]. In addition, Taverna *et al.* [41] demonstrated an association between risk of French type 1 (insulin-dependent) diabetic retinopathy patients and polymorphism of the vitamin D receptor.

Furthermore, Yokota *et al.* [42] reported an association between vitamin D receptor genotype and age of onset in juvenile Japanese patients with type 1 diabetes. Recently, Motohashi *et al.* [43] found an association between a VDR gene polymorphism and acute onset of type 1 diabetes, regardless of the presence or absence of islet-associated autoantibody. However, VDR gene polymorphisms in patients with type 1 diabetes do not have an effect on biochemical parameters of bone metabolism [44].

Vitamin D-binding protein (DBP) is essential for vitamin D cellular endocytosis and metabolism [45], thus variants of the DBP protein may affect the amount of active vitamin D on β cells and, subsequently, insulin

secretion. It has been found that the locus of the DBP gene was linked to plasma glucose and insulin concentrations in nondiabetic Pima Indians [46]. In a Hispanic-American/Anglo population of the San Luis Valley in Colorado, a variation in DBP is associated with elevated plasma glucose [47]. Genetic variants of the DBP have been reported to be associated with type 1 diabetes [48,49].

Vitamin D 1 α -hydroxylase (CYP1 α) is the key enzyme for both systemic and tissue levels of 1,25(OH)₂D₃ [50,51]. Induction of CYP1 α expression was found to be defective in macrophages of diabetic NOD mice [52]. Malecki *et al.* [53] reported the association of CYP 1 α gene and type 2 diabetes in a Polish population.

CYP27B1 (25-hydroxy-vitamin D₃-1 α -hydroxylase) enzyme catalyzes the 1 α -hydroxylation of the 25-hydroxy-vitamin D₃ to 1,25(OH)₂D₃, the most active form of vitamin D₃ metabolite. Interestingly, CYP27B1 polymorphisms variants have been reported to associate with type 1 diabetes mellitus in Germans [54].

Role of vitamin D in diabetes

Several studies in rats and humans [55,56] have demonstrated that vitamin D deficiency causes reduced insulin secretion, and that 1,25(OH)₂D₃ improves in β -cell function and consequently in glucose tolerance [57]. In vitamin D-deficient rats, glucose tolerance and insulin secretion were improved with 1,25(OH)₂D₃ treatment [58]. In gestational diabetes mellitus, Rudnicki and Molsted-Petersen [59] reported that the glucose level decreased from 5.6 to 4.8 mmol/L after intravenous treatment with 1,25(OH)₂D₃. This vitamin D also corrects glucose intolerance and normalizes insulin sensitivity in uremic patients [60,61].

The NOD mouse has been known as a model of human type 1 diabetes. Similar to the human disease, NOD mouse strain developed hyperglycemia as a result of a T-cell mediated autoimmune reaction against the insulin-producing β cells of the islets of Langerhans in the pancreas. Clinical disease is preceded by insulinitis, which is a basic histological lesion in the islet of Langerhans of the pancreas. Islets are invaded mainly by CD4⁺ and CD8⁺ T cells and also by monocytes [62]. The onset of insulinitis is observed at 20 to 40 days of age. The loss of glycemic control results in polydipsia, polyuria, and excessive weight loss if not treated with exogenous insulin, becoming 100% by 200 days. Mathieu *et al.* [63] reported that 1,25(OH)₂D₃ has been shown to reduce type 1 diabetes onset in NOD mice. No mouse (100%) showed insulinitis at 21 days of age. 1,25(OH)₂D₃ has shown partial protection, reduction to 42%, against insulinitis by 100 days of age. When 1,25(OH)₂D₃ treatment (on alternate days) was started at the age of 21 days and terminated at the age of 200 days or on the day of diabetes diagnosis [64], it reduced insulinitis incidence from 81% in the control group to 58% in the treated group. Diabetes incidence in female NOD mice at 200 days

was reduced to 8% in the $1,25(\text{OH})_2\text{D}_3$ -treated group versus 56% in the control group. Both $1,25(\text{OH})_2\text{D}_3$ and its nonhypercalcemic analogs, $1\alpha,25(\text{OH})_2\text{-20-epi-22-oxa-24,26,27-trishomo-vitamin D}$ (KH1060), have been shown to reduce type 1 diabetes onset in NOD mice [65]. However, Zella and DeLuca [5] found that $1,25(\text{OH})_2\text{D}_3$ does not offer complete protection against type 1 diabetes onset in NOD mice when administered every other day. They also showed that all NOD mice are completely resistant to type 1 diabetes by 200 days of age when a daily dose of 50-ng $1,25(\text{OH})_2\text{D}_3$ is administered orally through the diet from weaning. They suggested that oral administration of $1,25(\text{OH})_2\text{D}_3$ or preferably a nonhypercalcemia analog would be more clinically relevant for the prevention of type 1 diabetes in humans. Recently, Giulietti *et al.* [66] reported that vitamin D deficiency in early life might increase type 1 diabetes in NOD mice. They found, at 250 days, that 35% male and 66% female vitamin D-deficient mice were diabetic compared to 15 and 45% of the control mice. In the vitamin D-deficient mice, higher IL-1 expression was detected in islets. Thymus and lymph nodes also contained less CD4CD62L⁺ cells; a defect in this cytokine profile might trigger the diabetes.

In addition, Casteels *et al.* [67] reported that nonhypercalcemic analogs of $1,25(\text{OH})_2\text{D}_3$ administered to NOD mice when the autoimmune disease is already active can prevent clinical diabetes when this therapy is combined with a short induction course of an immunosuppressant such as Cyclosporin A (CsA).

Mechanism of vitamin D in type 1 diabetes mellitus

At the molecular level

To clarify the role of vitamin D in the regulation of the endocrine pancreas, some studies [68,69] suggested that vitamin and its metabolites act not only via the plasma calcium levels but also directly on the β cells. $1,25(\text{OH})_2\text{D}_3$ may influence both endocrine and exocrine pancreatic function [70]. The effects of $1,25(\text{OH})_2\text{D}_3$, a biologically active metabolite of vitamin D, and its analogs have been examined regarding binding to nuclear VDR (nVDR) and membrane VDR (mVDR), through which they might induce genomic and nongenomic responses respectively.

Kajikawa *et al.* [71] studied the effect of $1,25$ -dihydroxyluminesterol₃ [$1,25(\text{OH})_2$ lumisterol₃] – an analog of $1,25(\text{OH})_2\text{D}_3$ that is preferred for its nongenomic action through putative signal transduction by binding to mVDR [72] – on insulin release from rat pancreatic β cells. They found an insulinotropic effect of this vitamin analog with increasing intracellular Ca^{2+} concentration in pancreatic β cells through nongenomic signal transduction. There is also evidence that $1,25(\text{OH})_2\text{D}_3$ directly influences insulin secretion in the β cell through

a rise in intracellular-free calcium concentration via the nonselective calcium channel, rather than the calcium-dependent inositol 1,4,5-triphosphate receptor-mediated pathway [73,74]. $1,25(\text{OH})_2\text{D}_3$ also exerted a stimulating effect on insulin release via protein kinase A activation, but reduced the supranormal cyclic adenosine monophosphate (AMP) synthesis [75]. $1,25(\text{OH})_2\text{D}_3$ may provide supplementary calcium to the β cell by regulating the intracellular signaling processes involving phospholipids metabolism, protein kinase C induction, Ca^{2+} mobilization, and Ca^{2+} entry by Ca^{2+} channels [76] (Figure 1).

Norman *et al.* [55] reported the presence in the pancreas of a vitamin D-dependent calcium-binding protein and cytosol receptor for the hormonal form of vitamin D, $1,25$ -dihydroxyvitamin D₃, suggesting an important role of vitamin D in the endocrine functioning of the pancreas. Vitamin D-deficient rats were unable to respond to a glucose challenge by secreting appropriate amounts of insulin [55] since insulin release *in vitro* is dependent on acute change in plasma calcium [77]. Glucagon secretion is also calcium-dependent [78], but secretion of this hormone was unaffected by $1,25(\text{OH})_2\text{D}_3$ treatment [55]. The genomic actions of $1,25(\text{OH})_2\text{D}_3$ on β -cells of the endocrine pancreas have been reported. Calbindin-D_{28K}, a calcium-binding protein that is thought to act as a facilitator of calcium diffusion in intestine and kidney [79], is known to be regulated by vitamin D in these tissues. In cells transfected with Calbindin D_{28K} [80], there was a marked increase in the expression of insulin mRNA. In addition, Calbindin D_{28K} overexpression was also associated with an increase in insulin content and

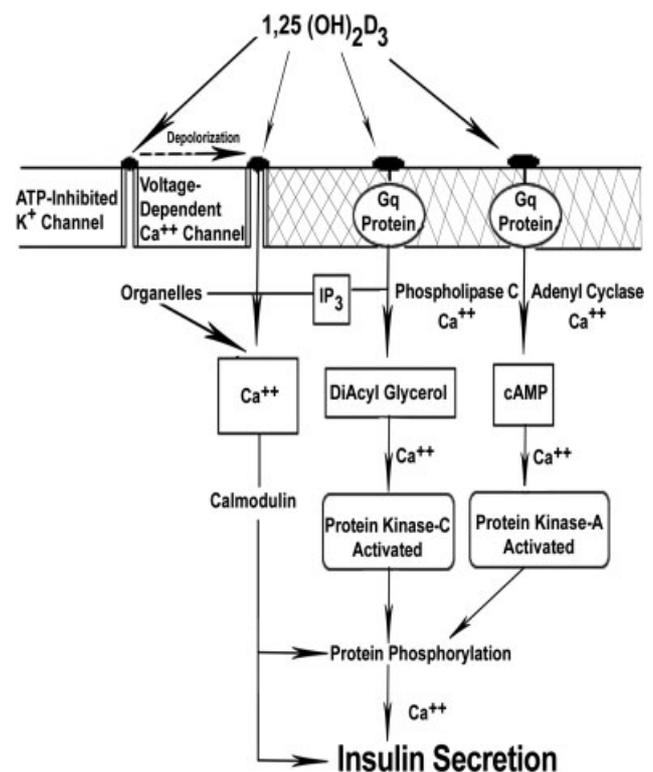


Figure 1. Schematic nongenomic model for $1,25(\text{OH})_2\text{D}_3$ effects on insulin secretion in β cells

release. In chicken, pancreatic Calbindin D_{28K} is altered with variations in vitamin D and mineral status [81]. 1,25(OH)₂D₃ activates the β cell insulin response to glucose *in vivo* after a delay of 3 to 20 h [57,82,83], or after 6-h *in vitro* [84], via an improvement of calcium handling occurring after a 4-h delay [84], increasing both Ca²⁺ entry by voltage-dependent channels and Ca²⁺ mobilization from Ca²⁺ stores [85,86]. Bourbon *et al.* [87] reported that *de novo* synthesis of numerous proteins is decreased during vitamin D₃ deficiency and is gradually restored by 1,25(OH)₂D₃ repletion in the islets of Langerhans of rats. The existence of variable delays for the actions of 1,25(OH)₂D₃ supports the hypothesis of a genomic action of the steroid on the biosynthesis of several β cell protein implicated in the different steps of the insulin excitation-secretion coupling. 1,25(OH)₂D₃ causes transcriptional activation of the human insulin receptor gene in U-937 human promonocytic cells [88]. Treatment with 1,25(OH)₂D₃ for 24 h increased, in a dose-dependent manner, the levels of the two major insulin receptor mRNA (11 and 8.5 Kb) present in U-937 human promonocytic cells [89], and preproinsulin (ppI) mRNA levels in 1,25(OH)₂D₃-replete rats increased at 8 and 24 h [90]. However, binding assay also indicated that 1,25(OH)₂D₃ increased the total insulin receptor number without altering receptor affinity [91].

Recently, Maestro *et al.* [92] reported vitamin D response elements (VDREs) in the insulin receptor (*hIR*) gene promoter that could account for the transcriptional induction of this gene by 1,25(OH)₂D₃ detected in U-937 cells. This locus could mediate cross talk between vitamin D and insulin-signaling pathways in U-937 cells. (Figure 2)

Vitamin D₃ and the immune system

The presence of the VDR in peripheral blood monocytes and activated T cells [1,2] has suggested a relationship between vitamin D and the immune system.

There were many reports on the immunological events that might trigger self-destruction of the pancreatic β -cells in type 1 diabetes. Progression of type 1 diabetes has been shown to involve infiltration into pancreatic islet cells by several types of immune cells including antigen-presenting cells (APCs – such as macrophages and dendritic cells), CD4⁺, and CD8⁺ T B cells, and B cells [93] (Figure 3).

Upon antigen stimulation, CD4⁺ cells differentiate into two distinct effectors populations, T helper 1 (Th1) and T helper 2 (Th2) cells. Their functions correlate well with their distinctive cytokines. Th1-type cytokines interleukin 2 (IL-2), interferon γ (IFN- γ), and tumor necrosis factor β (TNF- β), which activate cell-mediated immunity, that is, cytotoxic and inflammatory responses mediated by T cells, natural killer (NK) cells, and macrophages. Th2-type cytokines (IL-4, IL-5, IL-6, IL-9, IL-10, IL-13) activate humoral immunity, that is, antibody production by β -cells [94]. Insulinitis lesion is β -cell destructive when

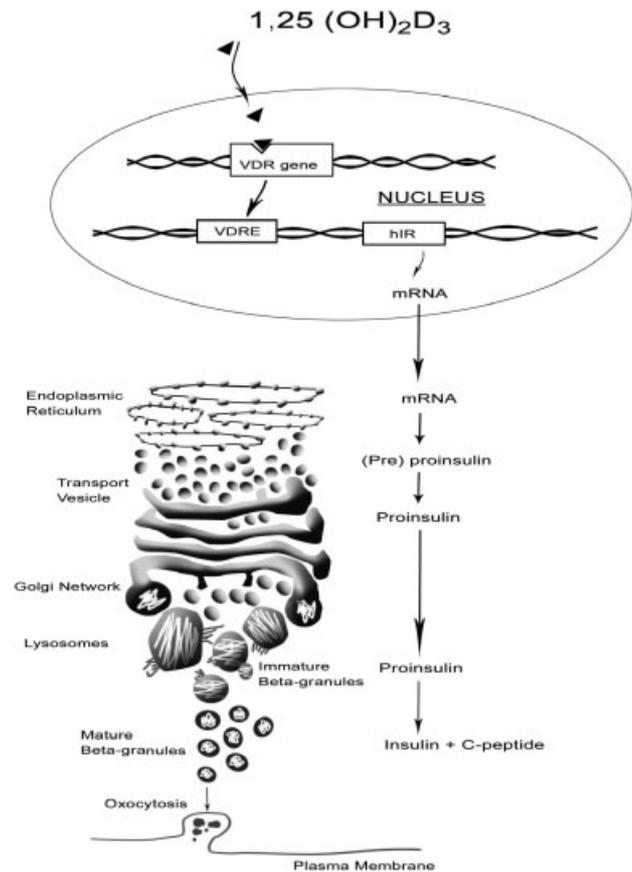


Figure 2. Schematic genomic model for 1,25(OH)₂D₃ effects on insulin secretion in β cells. The occupied vitamin D receptor (VDR) binds to the transcriptional domain of the insulin receptor (*hIR*) gene promoter, which is composed of a vitamin D response element (VDRE). Transcription induction by the VDR-complex increases the new insulin mRNA and protein. Newly synthesized proinsulin is formed in the rough endoplasmic reticulum (RER), then it is transferred to the Golgi network, where immature β -granules form. Proinsulin is targeted to the β -granule compartment, where it undergoes proteolytic conversion to insulin and C-peptide

Th1 cytokines produced by islet-infiltrating leukocytes dominate over Th2, and that insulinitis is benign when Th2 dominate and downregulate (suppress) Th1 cytokine production, thereby preventing β -cell destruction [94].

1,25(OH)₂D₃ treatment induces an autoantigen-specific 'protective' Th2 cell population not only at the site of the β -cell attack but also in the peripheral immune system [95]. This vitamin D also has a direct effect on naive CD4⁺ T cells to enhance the development of Th2 cells [96] in the absence of APC.

Dendritic cells (DCs) play a central role in regulating immune activation and response to self. DC maturation is central to the outcome of antigen presentation to T cells. Differentiation and maturation of DCs into potent APC are inhibited by physiological levels of 1,25(OH)₂D₃ and its analogs [97,98]. In NOD mice, 1,25(OH)₂D₃ analog treatment prevents DCs maturation, decreases liposaccharide-induced IL-12 and α -interferon production, enhances CD4⁺ CD25⁺ regulatory cells, arrests Th1 infiltration and progression of insulinitis, and

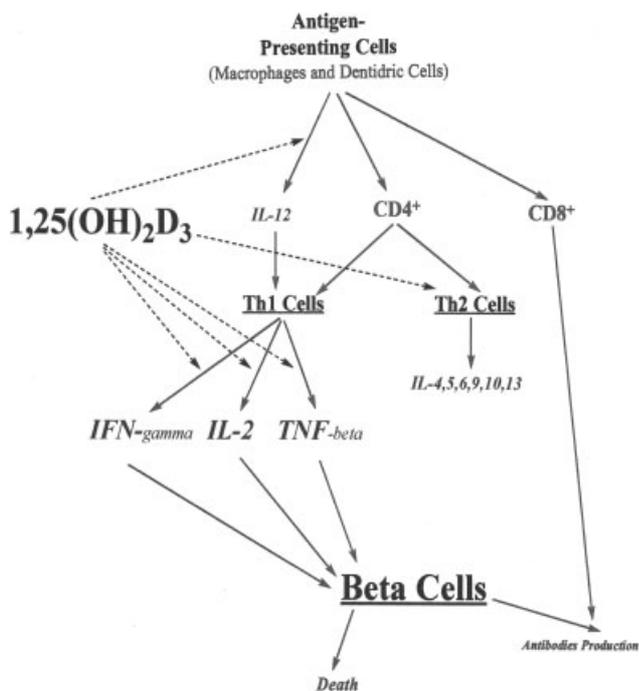


Figure 3. Role of $1,25(\text{OH})_2\text{D}_3$ in protecting β cells

inhibits diabetes development at nonhypercalcemic dose [25].

$1,25(\text{OH})_2\text{D}_3$ treatment induced inhibition of peripheral blood mononuclear cells (PBM) proliferation mediated through selective inhibition of IL-2 production [99]. Mathieu *et al.* [63,64] demonstrated a restoration of suppressor cell activity of $1,25(\text{OH})_2\text{D}_3$ and its analogs *in vitro* and *in vivo*.

Fujihira *et al.* [100] suggested that IL-12 may be an important regulator of effector cell development and activation in type 1 diabetes by neutralization of endogenous IL-12 with anti-IL-12 antibody that has been shown to ameliorate both insulinitis and diabetes in NOD mice. Since $1,25(\text{OH})_2\text{D}_3$ has been known to inhibit the production of IFN- γ , IL-2, and IL-12 [101], short-term treatment of adult NOD mice with an analog of $1,25(\text{OH})_2\text{D}_3$ inhibited IL-12 production, blocked pancreatic infiltration of Th1 cells, and arrested the progression of type 1 diabetes [102]. Therefore, vitamin D may be able to disrupt both the initiation and progression of the Th1-mediated pathogenesis of type 1 diabetes [65].

To investigate whether the restoration of the defective immune regulator system of the NOD mice is the only mechanism in the prevention of diabetes by $1,25(\text{OH})_2\text{D}_3$, Casteels *et al.* [103] tested if $1,25(\text{OH})_2\text{D}_3$ could prevent cyclophosphamide-induced diabetes, since diabetes occurring after cyclophosphamide injection is believed to be because of the elimination of suppressor cell. They found that NOD mice treated with $1,25(\text{OH})_2\text{D}_3$ from the time of weaning were clearly protected against diabetes induced by cyclophosphamide. They suggested that the protection against diabetes offered by $1,25(\text{OH})_2\text{D}_3$ may be independent of the presence of suppressor cells, and may involve increased apoptosis of

Th1 autoimmune effector. In addition, Gysemans *et al.* [104] reported that vitamin D_3 analogs (TX527) might have a role in preventing autoimmune diabetes recurrence after islet transplantation in spontaneously diabetic NOD mice. Mice treated with a combination of Interferon- β (IFN) and CsA showed no delay in autoimmune diabetes recurrence. However, 67% of mice treated with CsA combined with TX527 demonstrated normoglycemic blood levels. Interestingly, IFN- β in combination with TX527 is effective (100%) in inhibiting autoimmune diabetes recurrence.

Summary

The relationship between vitamin D and diabetes has been discussed in the literature. The potential for vitamin D supplement in the type 1 diabetes would be interested. However, $1,25(\text{OH})_2\text{D}_3$ enhances the insulin secretion. It does not participate in generating the new β -cells. Therefore, $1,25(\text{OH})_2\text{D}_3$ seems to have a role in the prevention of diabetes in early age and/or improving of diabetes rather than treating the disease. In addition, hypercalcemia is the most serious side effect of vitamin treatment, which may be lethal. Further investigation with nonhypercalcemic analogs of $1,25(\text{OH})_2\text{D}_3$ would be needed.

References

1. Bhalla AK, Amento EP, Clemens TL, *et al.* Specific high-affinity receptors for 1,25-dihydroxyvitamin D_3 in human peripheral blood mononuclear cells: Presence in monocytes and induction in T lymphocytes following activation. *J Clin Endocrinol Metab* 1983; **57**: 1308–1310.
2. Provedini DM, Tsoukas CD, Deftos LJ, Manolagas SC. 1,25-dihydroxyvitamin D_3 receptors in human leukocytes. *Science* 1983; **221**: 1181–1183.
3. Lemire JM, Archer DC. 1,25-dihydroxyvitamin D_3 prevents the *in vivo* induction of murine experimental autoimmune encephalomyelitis. *J Clin Invest* 1991; **87**: 1103–1107.
4. Cantorna MT, Hayes CE, DeLuca HF. 1,25-dihydroxycholecalciferol inhibits the progression of arthritis in murine models of models of human arthritis. *J Nutr* 1998; **128**: 68–72.
5. Zella JB, DeLuca HF. Vitamin D and autoimmune diabetes. *J Cell Biochem* 2003; **88**: 216–222.
6. Keen H, Ekoe JM. The geography of diabetes mellitus. *Br Med Bull* 1984; **40**: 359–365.
7. Diabetes Epidemiology Research International Group. Geographic patterns of childhood insulin-dependent diabetes mellitus. *Diabetes* 1988; **37**: 1113–1119.
8. Tuomilehto J, Rewers M, Reunanen A, *et al.* Increasing trend in type 1 (insulin-dependent) diabetes mellitus in childhood in Finland. Analysis of age, calendar time and birth cohort effects during 1965 to 1984. *Diabetologia* 1991; **34**: 282–287.
9. Padaiga Z, Tuomilehto J, Karvonen M, *et al.* Seasonal variation in the incidence of type 1 diabetes mellitus during 1983 to 1992 in the countries around the Baltic Sea. *Diabet Med* 1999; **16**: 736–743.
10. Ishii H, Suzuki H, Baba T, *et al.* Seasonal variation of glycemic control in type 2 diabetic patients. *Diabetes Care* 2001; **24**: 1503.
11. Muntoni S, Karvonen M, Muntoni S, Tuomilehto J. Seasonality in birth in patients with type 1 diabetes. *Lancet* 2002; **359**: 1246.
12. McKenna MJ. Differences in vitamin D status between countries in young adult and the elderly. *Am J Med* 1992; **93**: 69–77.

13. Webb AR, Kline L, Holick MF. Influence of season and latitude on the cutaneous synthesis of vitamin D₃: exposure to winter sunlight in Boston and Edmonton will not promote vitamin D₃ synthesis in human skin. *J Clin Endocrinol Metab* 1988; **67**: 373–378.
14. Fairney A, Fry J, Lipscomb A. The effect of darkness on vitamin D in adults. *Postgrad Med J* 1979; **55**: 248–250.
15. Stryd RP, Gilbertson TJ, Brunden MN. A seasonal variation study of 25-hydroxyvitamin D₃ serum levels in normal human. *J Clin Endocrinol Metab* 1979; **48**: 771–775.
16. Oliveri MB, Ladizesky M, Mautalen CA, *et al.* Seasonal variations of 25 hydroxyvitamin D and parathyroid hormone in Ushuaia (Argentina), the southernmost city of the world. *Bone Miner* 1993; **20**: 99–108.
17. Barger-Lux MJ, Heaney RP. Effects of above average summer sun exposure on serum 25-hydroxyvitamin D and calcium absorption. *J Clin Endocrinol Metab* 2002; **87**: 4952–4956.
18. Norris JM, Barriga K, Klingensmith G, *et al.* Timing of initial cereal exposure in infancy and risk of islet autoimmunity. *JAMA* 2003; **290**: 1713–1720.
19. Ziegler A, Schmid S, Huber D, *et al.* Early infant feeding and risk of developing type 1 diabetes-associated autoantibodies. *JAMA* 2003; **290**: 1721–1728.
20. Pileggi VJ, DeLuca HF, Steenbock H. The role of vitamin D and intestinal phytase in the prevention of rickets in rats on cereal diets. *Arch Biochem Biophys* 1955; **58**: 194–204.
21. Robertson I, Ford JA, McIntosh WB, Dunnigan MG. The role of cereals in the aetiology of nutritional rickets: the lesson of the Irish National Nutritional Survey 1943–1948. *Br J Nutr* 1981; **45**: 17–22.
22. Ford JA, McIntosh WB, Dunnigan MG. A possible relationship between high extraction cereal and rickets and osteomalacia. *Adv Exp Med Biol* 1977; **81**: 353–362.
23. Corazza GR, Di Sario A, Cecchetti L, *et al.* Influence of pattern of clinical presentation and of gluten-free diet on bone mass and metabolism in adult coeliac disease. *Bone* 1996; **18**: 525–530.
24. Dabadghao P, Bhatia E, Bhatia V, *et al.* Islet-cell antibodies in malnutrition-related diabetes mellitus from north India. *Diabetes Res Clin Pract* 1996; **34**: 73–78.
25. Wiyono P, Morimoto Y, Taniguchi H. The presence of islet-cell antibodies in malnutrition-related diabetes mellitus. *Diabetes Res Clin Pract* 1989; **6**: 75–78.
26. Gregori S, Giarratana N, Smiroldo S, *et al.* A 1 α ,25-dihydroxyvitamin D₃ analogs enhances regulatory T-cells and arrests autoimmune diabetes in NOD mice. *Diabetes* 2002; **51**: 1367–1374.
27. Baumgartl HL, Standl E, Schmidt-Gayk H, *et al.* Changes of vitamin D₃ serum concentrations at the onset of immune-mediated type 1 (insulin-dependent) diabetes mellitus. *Diabetes Res* 1991; **16**: 145–148.
28. The EURODIAB Substudy 2 Study Group. Vitamin supplement in early childhood and risk for type 1 (insulin-dependent) diabetes mellitus. *Diabetologia* 1999; **42**: 51–54.
29. Stene LC, Ulriksen J, Magnus P, Joner G. Use of code liver oil during pregnancy associated with lower risk of type 1 diabetes in the offspring. *Diabetologia* 2000; **43**: 1093–1098.
30. Hyponen E, Laara E, Reunanen A, *et al.* Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *Lancet* 2001; **358**: 1500–1503.
31. Lee S, Clark SA, Gill RK, Christakos S. 1,25-dihydroxyvitamin D₃ and pancreatic β -cell function: vitamin D receptors, gene expression and insulin secretion. *Endocrinology* 1994; **134**: 1602–1610.
32. Ortlepp JR, Metrikat J, Albrecht M, *et al.* The vitamin D receptor gene variant and physical activity predicts fasting glucose levels in healthy young men. *Diabet Med* 2003; **20**: 451–454.
33. McDermott ME, Ramachandran A, Ogunkolade BW, *et al.* Allelic variation in the vitamin D receptor influences susceptibility to IDDM in Indian Asians. *Diabetologia* 1997; **40**: 971–975.
34. Hauache OM, Lazaretti-Castro M, Andreoni S, *et al.* Vitamin D receptor gene polymorphism: correlation with bone mineral density in a Brazilian population with insulin-dependent diabetes mellitus. *Osteoporos Int* 1998; **8**: 204–210.
35. Chang TJ, Lei HH, Yeh JI, *et al.* Vitamin D receptor gene polymorphisms influence susceptibility to type 1 diabetes mellitus in the Taiwanese population. *Clin Endocrinol* 2000; **52**: 575–580.
36. Pani MA, Knapp M, Donner H, *et al.* Vitamin D receptor allele combinations influence genetic susceptibility to type 1 diabetes in Germans. *Diabetes* 2000; **49**: 504–507.
37. Ban Y, Taniyama M, Yanagawa T, *et al.* Vitamin D receptor initiation codon polymorphism influences genetic susceptibility to type 1 diabetes mellitus in the Japanese population. *BMC Med Genet* 2001; **2**: 7–12.
38. Ogunkolade BW, Boucher BJ, Prahil JM, *et al.* Vitamin D receptor (VDR) mRNA and VDR protein levels in relation to vitamin D status, insulin secretory capacity, and VDR genotype in Bangladeshi Asians. *Diabetes* 2002; **51**: 2294–2312.
39. Guja C, Marshall S, Welsh K, *et al.* The study of CTLA-4 and vitamin D receptor polymorphisms in the Romanian type 1 diabetes population. *J Cell Mol Med* 2002; **6**: 75–81.
40. Skrabic V, Zemunik T, Situm M, Terzic J. Vitamin D receptor polymorphisms and susceptibility to type 1 diabetes in the Dalmatian population. *Diabetes Res Clin Pract* 2003; **59**: 31–35.
41. Taverna MJ, Sola A, Guyot-Argenton C, *et al.* Taq I polymorphism of the vitamin D receptor and risk of severe diabetic retinopathy. *Diabetologia* 2002; **45**: 436–442.
42. Yokota I, Satomura S, Kitamura S, *et al.* Association between vitamin D receptor genotype and age of onset in juvenile Japanese patients with type 1 diabetes. *Diabetes Care* 2002; **25**: 1244.
43. Motohashi Y, Yamada S, Yanagawa T, *et al.* Vitamin D receptor gene polymorphism affects onset pattern of type 1 diabetes. *J Clin Endocrinol Metab* 2003; **88**: 3137–3140.
44. Fassbender WJ, Goertz B, Weismuller K, *et al.* VDR gene polymorphisms are overrepresented in German patients with type 1 diabetes compared to healthy controls without effect on biochemical parameters of bone metabolism. *Horm Metab Res* 2002; **34**: 330–337.
45. Nykjaer A, Dragun D, Walther D, *et al.* An endocytic pathway essential for renal uptake and activation of the steroid 25-(OH) Vitamin D₃. *Cell* 1999; **96**: 507–515.
46. Pratley RE, Thompson DB, Prochazka M, *et al.* An autosomal genomic scan for loci linked to prediabetic phenotype in Pima Indians. *J Clin Invest* 1998; **101**: 1757–1764.
47. Iyengar S, Hamman RF, Marshall JA, Majumder PP, Ferrell RE. On the role of vitamin D binding globulin in glucose homeostasis: results from the San Luis Valley Diabetes Study. *Genet Epidemiol* 1989; **6**: 691–698.
48. Hodge SE, Anderson CE, Neiswanger K, *et al.* Association studies between Type 1 (insulin-dependent) diabetes and 27 genetic markers: lack of association between type 1 diabetes and Kidd blood group. *Diabetologia* 1983; **25**: 343–347.
49. Ongagna JC, Kaltenbacher MC, Sapin R, Pinget M, Belcourt A. The HLA-DQB alleles and amino acid variants of the vitamin D-binding protein in diabetic patients in Alsace. *Clin Biochem* 2001; **34**: 59–63.
50. Hewison M, Zehnder D, Bland R, Stewart PM. 1 α -Hydroxylase and the action of vitamin D. *J Mol Endocrinol* 2000; **25**: 141–148.
51. Miller WL, Portale AA. Vitamin D 1 α -Hydroxylase. *Trends Endocrinol Metab* 2000; **11**: 315–319.
52. Overbergh L, Decallonne B, Valckx D, *et al.* Identification and immune regulation of 25-hydroxyvitamin D-1 α -hydroxylase in murine macrophages. *Clin Exp Immunol* 2000; **120**: 139–146.
53. Malecki MT, Klupa T, Wolkow P, *et al.* Association study of the vitamin D: 1 α -hydroxylase (CYP1 α) gene and type 2 diabetes mellitus in a Polish population. *Diabetes Metab* 2003; **29**: 119–124.
54. Lopez ER, Regulla K, Pani MA, *et al.* CYP27B1 polymorphisms variants are associated with type 1 diabetes mellitus in Germans. *J Steroid Biochem Mol Biol* 2004; **89–90**: 155–157.
55. Norman AW, Frankel BJ, Heldt AM, Grodsky GM. Vitamin D deficiency inhibits pancreatic secretions of insulin. *Science* 1980; **209**: 823–825.
56. Boucher BJ, Mannan N, Noonan K, *et al.* Glucose intolerance and impairment of insulin secretion in relation to vitamin D deficiency in East London Asians. *Diabetologia* 1995; **38**: 1239–1245.
57. Kumar S, Davies M, Zakaria Y, *et al.* Improvement in glucose tolerance and β -cell function in a patient with vitamin D deficiency during treatment with vitamin D. *Postgrad Med J* 1994; **70**: 440–443.
58. Cade C, Norman AW. Rapid normalization/stimulation by 1,25-dihydroxyvitamin D₃ on insulin secretion and glucose

- tolerance in the vitamin D-deficient rat. *Endocrinology* 1987; **120**: 1490–1497.
59. Rudnicki PM, Molsted-Pedersen L. Effect of 1,25-dihydroxycholecalciferol on glucose metabolism in gestational diabetes mellitus. *Diabetologia* 1997; **40**: 40–44.
 60. Mak RH. Intravenous 1,25 dihydroxycholecalciferol corrects glucose intolerance in hemodialysis patients. *Kidney Int* 1992; **41**: 1049–1054.
 61. Kautzky-Willer A, Pacini G, Barnas U, et al. Intravenous calcitriol normalizes insulin sensitivity in uremic patients. *Kidney Int* 1995; **47**: 200–206.
 62. Lampeter F, Signore A, Gale E, et al. Lessons from the NOD mouse for the pathogenesis and immunotherapy of human type 1 (insulin-dependent) diabetes mellitus. *Diabetologia* 1989; **32**: 703–708.
 63. Mathieu C, Laureys J, Sobis H, et al. 1,25-dihydroxyvitamin D₃ prevents insulinitis in NOD mice. *Diabetes* 1992; **41**: 1491–1495.
 64. Mathieu C, Waer M, Laureys J, et al. Prevention of autoimmune diabetes in NOD mice by 1,25-dihydroxyvitamin D₃. *Diabetologia* 1994; **37**: 552–558.
 65. Mathieu C, Waer M, Casteels K, et al. Prevention of type 1 diabetes in NOD mice by nonhypercalcemic doses of a new structural analog of 1,25-dihydroxyvitamin D₃, KH1060. *Endocrinology* 1995; **136**: 866–872.
 66. Giulietti A, Gysemans C, Stoffels K, et al. Vitamin D deficiency in early life accelerates Type 1 diabetes in non-obese diabetic mice. *Diabetologia* 2004; **47**: 451–462.
 67. Casteels KM, Mathieu C, Waer M, et al. Prevention of type 1 diabetes in nonobese diabetic mice by late intervention with nonhypercalcemic analogs of 1,25-dihydroxyvitamin D₃ in combination with a short induction course of cyclosporin A. *Endocrinology* 1998; **139**: 95–102.
 68. Tanaka Y, Seino Y, Ishida M, et al. Effect of vitamin D₃ on the pancreatic secretion of insulin and somatostatin. *Acta Endocrinol* 1984; **105**: 528–533.
 69. Kadowaki S, Norman AW. Dietary vitamin D is essential for normal insulin secretion from the perfused rat pancreas. *J Clin Invest* 1984; **73**: 759–766.
 70. Johnson JA, Grande JP, Roche PC, Kumar R. Immunohistochemical localization of the 1,25(OH)₂D₃ receptor and calbindin D_{28K} in human and rat pancreas. *Am J Physiol* 1994; **267**: E356–E360.
 71. Kajikawa M, Ishida H, Fujimoto S, et al. An insulinotropic effect of vitamin D analog with increasing intracellular Ca²⁺ concentration in pancreatic β-cells through nongenomic signal transduction. *Endocrinology* 1999; **140**: 4706–4712.
 72. Dormanen MC, Bishop JE, Hammond MW, et al. Nonnuclear effects of the steroid hormone 1,25-dihydroxyvitamin D₃ analogs are able to functionally differentiate between nuclear and membrane receptors. *Biochem Biophys Res Commun* 1994; **201**: 394–401.
 73. Wollheim CB, Blondel B, Trueheart PA, et al. Calcium-induced insulin release in monolayer culture of the endocrine pancreas. *J Biol Chem* 1975; **250**: 1354–1360.
 74. Sergeev IN, Rhoten WB. 1,25-dihydroxyvitamin D₃ evokes oscillations of intracellular calcium in a pancreatic β-cell line. *Endocrinology* 1995; **136**: 2852–2861.
 75. Bourlon PM, Faure-Dussert AG, Billaudel BJ. Modulatory role of 1,25-dihydroxyvitamin D₃ on pancreatic islet insulin release via the cyclic AMP pathway in the rat. *Br J Pharmacol* 1997; **121**: 751–758.
 76. Billaudel BJ, Bourlon PM, Sutter BC, Faure-Dussert AG. Regulatory effect of 1,25-dihydroxyvitamin D₃ on insulin release and calcium handling via the phospholipid pathway in islets from vitamin D-deficient rats. *J Endocrinol Invest* 1995; **18**: 673–682.
 77. Curry DL, Bennett LL, Grodsky GM. Dynamics of insulin secretion by the perfused rat pancreas. *Endocrinology* 1968; **83**: 572–584.
 78. Lundquist I, Fanska R, Grodsky GM. Interaction of calcium and glucose on glucagon secretion. *Endocrinology* 1976; **99**: 1304–1312.
 79. Christakos S, Gabrielides C, Rhoten WB. Vitamin D-dependent calcium binding proteins: chemistry, distribution, functional considerations, and molecular biology. *Endocr Rev* 1989; **10**: 3–26.
 80. Reddy D, Pollock AS, Clark SA, et al. Transfection and overexpression of the calcium binding protein calbindin-D28K results in a stimulatory effect on insulin synthesis in a rat β cell line (RIN 1046-38). *Proc Natl Acad Sci U S A* 1997; **94**: 1961–1966.
 81. Kadowaki S, Norman AW. Pancreatic vitamin D-dependent calcium binding protein: biochemical properties and response to vitamin D. *Arch Biochem Biophys* 1984; **233**: 228–236.
 82. Ishida H, Seino Y, Sieno S, et al. Effect of 1,25-dihydroxyvitamin D₃ on pancreatic B and D cell function. *Life Sci* 1983; **33**: 1779–1786.
 83. Kadowaki S, Norman AW. Time course study of insulin secretion after 1,25-dihydroxyvitamin D₃ administration. *Endocrinology* 1985; **117**: 1765–1771.
 84. Billaudel BJ, Faure AG, Sutter BC. Effect of 1,25-dihydroxyvitamin D₃ on isolated islets from vitamin D₃-deprived rats. *Am J Physiol* 1990; **258**: E643–E648.
 85. Faure AG, Sutter BC, Billaudel BJ. Is 1,25-dihydroxyvitamin D₃ the specific vitamin D₃ metabolite active on insulin release and calcium handling by islets from vitamin D₃-deprived rats? *Diabetes Metab* 1991; **17**: 271–278.
 86. Billaudel BJ, Delbancut AP, Sutter BC, Faure AG. Stimulatory effect of 1,25-dihydroxyvitamin D₃ on calcium handling and insulin secretion by islets from vitamin D₃-deficient rats. *Steroids* 1993; **58**: 335–341.
 87. Bourlon PM, Faure-Dussert AG, Billaudel BJ. The *de novo* synthesis of numerous proteins is decreased during vitamin D₃ deficiency and is gradually restored by 1,25-dihydroxyvitamin D₃ repletion in the islets of Langerhans of rats. *J Endocrinol* 1999; **162**: 101–109.
 88. Maestro B, Molero S, Bajo S, et al. Transcriptional activation of the human insulin receptor gene by 1,25-dihydroxyvitamin D₃. *Cell Biochem Funct* 2002; **20**: 227–232.
 89. Maestro B, Campion J, Davila N, Calle C. Stimulation by 1,25-dihydroxyvitamin D₃ of insulin receptor expression and insulin responsiveness for glucose transport in U-937 human promonocytic cells. *Endocr J* 2000; **47**: 383–391.
 90. Ozone K, Seino Y, Yano H, et al. 1,25-dihydroxyvitamin D₃ enhances the effects of refeeding on steady state preproinsulin messenger ribonucleic acid levels in rats. *Endocrinology* 1990; **126**: 2041–2045.
 91. Leal MA, Aller P, Mas A, Calle C. The effect of 1,25-dihydroxyvitamin D₃ on insulin binding, insulin receptor mRNA levels, and isotope RNA pattern in U-937 human promonocytic cells. *Exp Cell Res* 1995; **217**: 189–194.
 92. Maestro B, Davila N, Carranza C, Calle C. Identification of a vitamin D response element in the human insulin receptor gene promoter. *J Steroid Biochem Mol Biol* 2003; **84**: 223–230.
 93. Benoist C, Mathis D. Cell death mediators in autoimmune diabetes—No shortage of suspects. *Cell* 1997; **89**: 1–3.
 94. Rabinovitch A. An update on cytokines in the pathogenesis of insulin-dependent diabetes mellitus. *Diabetes Metab Rev* 1998; **14**: 129–151.
 95. Overbergh L, Decallonne B, Waer M, et al. 1,25-dihydroxyvitamin D₃ induces an autoantigen-specific T-helper 1/T-helper 2 immune shift in NOD mice immunized with GAD65 (p524-543). *Diabetes* 2000; **49**: 1301–1307.
 96. Boonstra A, Barrat FJ, Crain C, et al. 1,25-dihydroxyvitamin D₃ has a direct effect on naive CD4⁺ T cells to enhance the development of Th2 cells. *J Immunol* 2001; **167**: 4974–4980.
 97. Piemonti L, Monti P, Sironi M, et al. Vitamin D₃ affects differentiation, maturation, and function of human monocyte-derived dendritic cells. *J Immunol* 2000; **164**: 4443–4451.
 98. Griffin MD, Luts W, Phan VA, et al. Dendritic cell modulation by 1,25-dihydroxyvitamin D₃ and its analogs: a vitamin D receptor-dependent pathway that promotes a persistent state of immaturity *in vitro* and *in vivo*. *Proc Natl Acad Sci U S A* 2001; **98**: 6800–6805.
 99. Manlagas SC, Provvedini DM, Murray EJ, et al. The antiproliferative effect of calcitriol on human peripheral blood mononuclear cells. *J Clin Endocrinol Metab* 1986; **63**: 394–400.
 100. Fujihira K, Nagata M, Moriyama H, et al. Suppression and acceleration of autoimmune diabetes by neutralization of endogenous interleukin-12 in NOD mice. *Diabetes* 2002; **51**: 1367–1374.
 101. Lemire JM. Immunomodulatory actions of 1,25-dihydroxyvitamin D₃. *J Steroid Biochem Mol Biol* 1995; **53**: 599–602.
 102. Castano L, Eisenbarth G. Type-1 diabetes: a chronic autoimmune disease of human, mouse and rat. *Annu Rev Immunol* 1990; **8**: 647–679.

103. Casteels KM, Waer M, Bouillon R, *et al.* 1,25-dihydroxyvitamin D₃ restores sensitivity to cyclophosphamide-induced apoptosis in non-obese diabetic (NOD) mice and protects against diabetes. *Clin Exp Immunol* 1998; **112**: 181–187.
104. Gysemans C, van Etten E, Overbergh L, *et al.* Treatment of autoimmune diabetes recurrence in non-obese diabetic mice by mouse interferon- β in combination with an analogue of 1 α ,25-dihydroxyvitamin-D₃. *Clin Exp Immunol* 2002; **128**: 213–220.