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## Serum uric acid and multiple sclerosis

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**Abstract** Several studies indicate that patients with multiple sclerosis (MS) have low serum levels of the endogenous antioxidant uric acid (UA), although it has not been established whether UA is primarily deficient or secondarily reduced due to its peroxynitrite scavenging activity. We measured serum urate levels in 124 MS patients and 124 age- and sex-matched controls with other neurological diseases. In addition, we compared UA levels when MS patients were stratified according to disease activity (by means of clinical examination and MRI), duration, disability and course. MS patients had significantly lower serum urate levels than controls ( $p=0.001$ ). However, UA levels did not significantly correlate with disease activity, duration, disability or course. Our study favors the view that reduced UA in MS is a primary, constitutive loss of protection against oxidative agents, which deserves further pathogenetic elucidation aimed at future therapeutic strategies.

**Key words** Multiple sclerosis • Uric acid • Nitric oxide • Peroxynitrite • Pathogenesis

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### Introduction

In rodent experimental autoimmune encephalomyelitis (EAE), the production of the free radical nitric oxide (NO) strongly correlated with the clinical severity of the disease [1]. The combination of NO with superoxide ( $O_2^-$ ) during an inflammatory response leads to the formation of peroxynitrite (PN). By virtue of its potent oxidant activity, PN is believed to be responsible for the majority of damage to the central nervous system (CNS) attributed to NO [2].

Since observations on EAE may apply to multiple sclerosis (MS) one could expect that levels of uric acid (UA), a natural scavenger of PN able to penetrate and preserve the blood-brain barrier integrity [3], may be abnormal in MS patients. Regardless of clinical status, MS patients have significantly lower serum UA levels than controls [4, 5]; these studies demonstrated a protective role of UA in EAE, and showed a beneficial function of raising serum UA levels in MS patients [4, 5]. Since gout prevalence in MS is 15-times less than that estimated for the general population [1], inadequate protection against PN-mediated CNS toxicity may be a primary, constitutive MS-related defect. One study indicated that UA reduction is strictly linked to clinical and MRI activity in MS patients [6]. It is not known whether UA reduction is primarily linked to MS or simply represents an epiphenomenon related to its PN scavenging activity, or both.

In the light of such considerations, we conducted a hospitalized-based study aimed at determining possible differences in serum UA levels in relation to MS clinical status.

### Patients and methods

#### Patients and controls

All subjects gave informed consent prior to their inclusion in the study.

A total of 124 in-patients (39 men) diagnosed as having probable or definite MS according to the criteria of Poser et al. [7] were randomly selected from the in-patient register of our Neurology

Clinic. Reasons for hospitalization were diagnostic purposes in patients with stable disease and therapeutic purposes in patients with clinically active disease (defined as the development within the previous two weeks of new neurological symptoms or signs attributable to demyelination). In a subgroup of 21 patients, 13 with active clinical disease and 8 with inactive disease, standard magnetic resonance imaging (MRI) of the brain (1.5 T) with T1-weighted sequences 15 minutes after injection of Gd-DTPA, was performed in the period from 2 days prior to serum UA determination to 2 days after.

As control population, we recruited 124 age- and sex- matched in-patients with other neurological diseases (OND) Clinical diagnosis was: syncope (19 patients), radiculopathy (16), trauma (14), neurosis (13), tumor (11), polyneuropathy (11), vertigo (13), myasthenia gravis (9), seizure (8), stroke (5), and myopathy (5).

Exclusion criteria for MS and OND patients were treatment with drugs that could increase serum levels of uric acid (e.g. steroids, acetylsalicylic acid and thiazide-type diuretics [8]) or reduce them (e.g. ibuprofen) [9]. Clinical or laboratory evidence of diabetes mellitus and renal failure were also ruled out in both groups. About 6% of controls had seizures. In general, increased urate in the postictal period may be observed and several drugs used in epilepsy may change levels of urate (e.g. carbamazepine and valproic acid may decrease whereas primidone may increase UA) [10]. Epileptic patients included in the study were not in the postictal period.

Blood samples were obtained before breakfast 5 days after patient's first hospitalization. Serum UA levels were determined by blinded technicians at the laboratory of Civic Hospital in Sassari, using a commercially available kit and following the manufacturer's instructions (Olympus Diagnostica, Hamburg, Germany). Normal ranges of UA, according to the Civic Hospital laboratory standardization, were 3.4–7.0 mg/dl. The levels of UA were compared when patients were stratified according to disability (scored on Kurtzke's expanded disability status scale, EDSS), disease duration, clinical activity, disease course and activity on MR images. Significance was calculated using the *t*-test and *p* value set at <0.05.

## Results

The mean UA level in MS patients was significantly lower than that in OND patients (Table 1). Mean urate levels were higher in men than in women in both groups. Among MS patients, urate levels were lower in patients with long duration (>1 year), more disabling disease (EDSS>3.5), active disease and secondary progressive course, although these differences were not statistically significant (Table 2).

**Table 1** Serum uric acid (UA) levels in hospitalized patients with multiple sclerosis (MS) and in matched control patients with other neurological diseases (OND)

		n	Age, years <sup>a</sup>	UA, mg/dl <sup>b</sup>	<i>t</i> -test
Female	MS	85	33.3 (16-61)	3.8 (1.1)	<i>p</i> =0.0004
	OND	84	33.5 (17-62)	4.2 (1.1)	
Male	MS	39	32.1 (18-50)	4.7 (1.3)	<i>p</i> =0.03
	OND	40	33.3 (19-56)	6.1 (1.9)	
Total	MS	124	33.1 (16-61)	4.1 (1.1)	<i>p</i> =0.001
	OND	124	33.5 (17-62)	4.7 (1.7)	

<sup>a</sup>Values are mean (range)

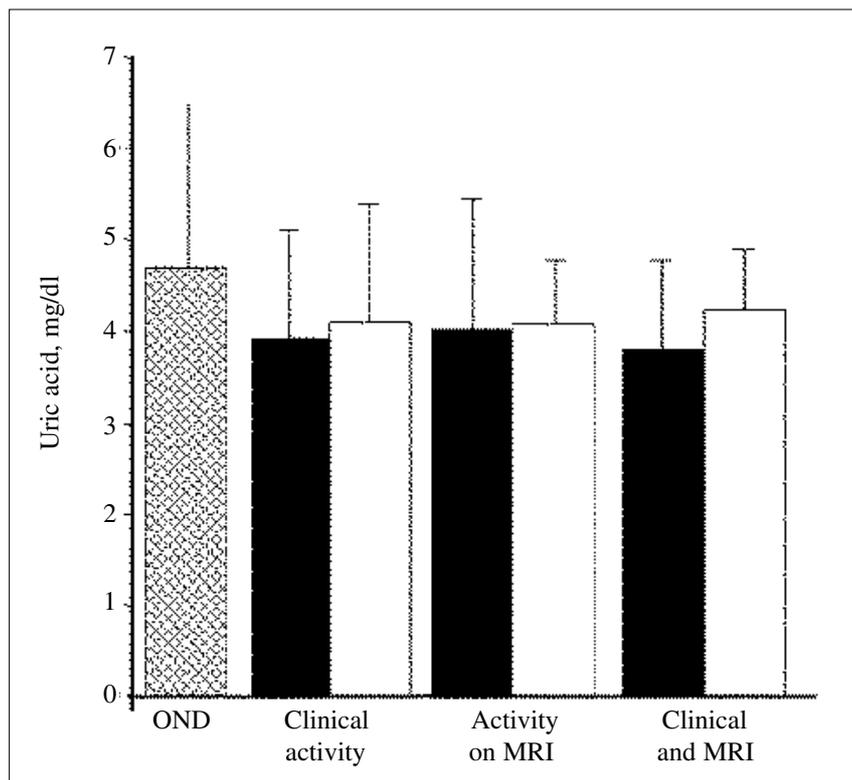
<sup>b</sup>Values are mean (SD)

**Table 2** Serum uric acid (UA) levels in 124 patients with multiple sclerosis (MS). Values of UA are mean (SD). No differences between groups are significant

	Patients, n	UA, mg/dl
Disease duration		
<1 year	41	4.1 (1.6)
≥1 year	83	4.1 (1.0)
Disease course		
Relapsing-remitting	79	4.1 (1.3)
Secondary progressive	45	3.9 (1.1)
Disability score		
EDSS <3.5	81	4.1 (1.3)
EDSS ≥ 3.5	43	4.0 (1.2)
Clinical activity		
Active	68	3.9 (1.2)
Inactive	56	4.1 (1.3)
Activity on MR images <sup>a</sup>		
Active	13	4.0 (1.5)
Inactive	8	4.1 (0.7)

EDSS, expanded disability status scale; MR, magnetic resonance

<sup>a</sup>In 21 patients



**Fig. 1** Serum levels of uric acid (UA) in 124 patients with multiple sclerosis (MS) and in 124 matched subjects with other neurological diseases (OND). *Black bars*, active disease; *white bars*, inactive disease

In the subgroup of 21 patients who underwent brain MRI, 13 (62%) patients, including 9 with clinically active disease and 4 with inactive disease, had one or more Gd-DTPA-enhancing lesions. The highest mean UA level (4.2 mg/dl) was found in the four clinically and MRI inactive patients whereas the lowest (3.8 mg/dl) was observed in the 9 clinically and MRI active ones (Fig. 1). However, these differences were not significant.

## Discussion

Among the toxic effects of PN on CNS tissues [11], lipid peroxidation [12], release of oxygen species and tyrosine nitration [13–15], and DNA strand breakage or mutation virtually leading to apoptosis of resident cells [13,16] are possibly the most relevant ones. A role of PN in the pathogenesis of MS is sustained by the detection of nitrotyrosinase residues on cell debris in MS plaques [16], anti-nitrotyrosine antibodies in MS sera [17], lipid peroxidation [18] and oxidized lipids and proteins found by means of MS lesion microspectroscopy [19]. However, myelin vacuolation and primary acute axonopathy are found in mice as a UA-treatable intrathecal PN effect, even in the absence of inflammation [20].

High levels of NO and metabolites (nitrite and nitrate) are found in the cerebrospinal fluid (CSF) of MS patients during an exacerbation [21, 22] despite some controversies

[23]. Urinary nitrite and nitrate levels are increased in early and relapsing-remitting MS but not in patients with progressive disease, thus confirming a participation of NO in modulating tissue damage [24] and in amplifying the effector stage of the immune attack against myelin [25, 26].

Induction of NO synthase (NOS) in CNS is commonly associated with cells of the monocyte lineage [27, 28] and with dendritic cells [29]. Interferon (IFN)- $\gamma$  is a major contributor to NOS induction [30], while IFN- $\gamma$  showed inhibitory effects on the IFN- $\beta$ -induced NOS expression in astrocytes [31]. When treated with the NOS inhibitors aminoguanidine [32] and lovastatin [33] and with an antisense oligodeoxynucleotide complementary to NOS [34], mice failed to develop EAE. Nonetheless, there is also evidence favoring the view that NO has a protective role in the CNS: (i) NOS knock-out mice had a greater incidence of EAE and higher clinical score than wild-type mice [35], (ii) locally activated dendritic cell-derived NO promotes apoptosis of auto-reactive T cells in EAE, therefore acting as a protective factor during the course of a T cell-mediated immune attack [29] and (iii) macrophages excrete large amounts of UA during active phagocytosis [36]. Additionally, a recent study suggested that specific inhibition of NO production in vivo rendered glucocorticoid receptor knock-out mice susceptible to EAE [37], thus indicating a complex, still debated contribution of NO in CNS tissue damage or repair. Finally, a genetic study in Swedish multiplex MS families clearly excluded NOS as a MS susceptibility gene region in this population [38].

Increased levels of UA in the spinal cord of rats during active EAE have been reported [39], and UA was increased within the MS plaque, reduced in adjacent white matter and even lowered in distant white matter [40]. Consistently, healthy monozygotic twins discordant for MS have higher serum levels of UA than the affected twin [41]. However, though the number of relapses showed a decremental rate, UA serum levels monitored during the course of IFN- $\beta$  treatment for MS did not change over time [42].

Serial observations [1, 4, 5] indicated that low UA levels are possibly a primary feature of MS patients. Instead, another study suggested that UA levels serve as a marker of MS activity, being inversely correlated with disease activity and duration [6]. In the present study, we obtained confirmatory evidence for an association of low serum UA levels with MS, even though some control neurological diseases (such as epilepsy and stroke) were potentially related to hyperuricemia. Drulovic and colleagues [6] found confirmatory evidence that patients with active relapsing-remitting and secondary-progressive forms of MS have significantly lower UA levels than controls. On the other hand, in the overall group (including MS patients with primary progressive and clinically inactive forms of MS) serum UA levels did not differ significantly from those of controls, although a strong tendency was evident ( $p=0.068$ ). It may be observed that the two study populations (MS and OND) of Drulovic et al. [6] differed in terms of numerosity (240 and 104, respectively) and that about 30% of their control patients had seizures. Since carbamazepine and valproic acid can decrease serum UA levels [10], they may possibly have lowered the mean UA level in the OND population.

In our study, comparison of UA levels in patients stratified according to MS course and disability showed no significant difference. This observation may favor the view that, although a concomitant disease activity-related effect is present, low UA levels might represent a primary "MS-specific" deficiency. In fact, despite the fact that the UA level was lower in MRI- and clinically-documented active patients, therefore partially confirming Drulovic et al's [6] results, a general mean UA decrease is evident also in clinically and MRI inactive MS patients as compared to OND.

Therefore, the question whether reduced UA level in MS is a primary deficit or an epiphenomenon related to its oxidation by PN and free radicals remains open, although the two alternative hypotheses are not mutually exclusive and concomitance of either facts is likely. In the latest metabolic stage, UA is transformed into allantoin which does not have PN scavenging activity. In other pathological conditions, such as myocardial infarction [43] and chronic lung disease of preterm infants [44] or during muscle metabolic stress induced by intense exercise [45], serum allantoin is a useful early predictor of the subsequent free radicals generation. To our knowledge, no study has assessed allantoin levels in MS patients so as to determine whether UA is primarily deficient or secondarily reduced by virtue of its protective role against

oxidant compounds. Investigations aimed at determining such an in vivo surrogate marker of free radical production in MS are clinically relevant. In fact, although the therapeutic value of raising UA levels in already established MS has been indicated [5, 46, 47], further pathogenetic investigations are needed so as to propose antioxidant prevention treatments in individuals at high-risk of developing MS (e.g. unaffected monozygotic twins, first-degree relatives of MS patients and individuals residing in areas at high risk for MS) and before MS clinical appearance.

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**Sommario** Numerosi lavori indicano come nei pazienti con sclerosi multipla (SM) vi siano ridotti livelli sierici dell'antiossidante endogeno acido urico (AU), sebbene non sia chiaramente stabilito se l'AU sia primariamente deficitario o secondariamente ridotto in seguito all'attività di scavenger del perossinitrito. Il nostro studio è volto a fornire evidenze in favore dell'una o l'altra ipotesi. Sono stati comparati i livelli medi ematici di AU da 124 pazienti con SM e 124 controlli neurologici appaiati. Inoltre, i livelli di AU dei pazienti con SM sono stati stratificati in base alle principali caratteristiche cliniche quali l'attività di malattia (clinica e di RM), la durata, la disabilità e il tipo di decorso. I dati indicano che i pazienti con SM presentano livelli ematici di AU significativamente ridotti rispetto a controlli con altre patologie neurologiche ( $p=0.001$ ). Tuttavia, i livelli di AU non correlano significativamente con il grado di attività, la durata, la disabilità, né col tipo di decorso della malattia. I risultati del nostro studio sembrano favorire l'ipotesi che la riduzione dell'AU nella SM costituisca un deficit primario di protezione contro gli agenti ossidanti. Tale ipotesi necessita di ulteriori conferme e delucidazioni al fine di intraprendere possibili nuove strategie terapeutiche.

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## References

1. Hooper DC, Bagasra O, Marini JC et al (1997) Prevention of experimental allergic encephalomyelitis by targeting nitric oxide and peroxynitrite: implications for the treatment of multiple sclerosis. *Proc Natl Acad Sci USA* 94:2528–2533
2. Van der Veen RC, Hinton DR, Incardonna F, Hofman FM (1997) Extensive peroxynitrite activity during progressive stages of central nervous system inflammation. *J Neuroimmunol* 77:1–7
3. Kean RB, Spitsin SV, Mikheeva T et al (2000) The peroxynitrite scavenger uric acid prevents inflammatory cell invasion into the central nervous system in experimental allergic

- encephalomyelitis through maintenance of blood-central nervous system barrier integrity. *J Immunol* 165:6511–6518
4. Hooper DC, Spitsin S, Kean RB, et al (1998) Uric acid, a natural scavenger of peroxynitrite, in experimental allergic encephalomyelitis and multiple sclerosis. *Proc Natl Acad Sci USA* 95:675–680
  5. Koprowski H, Spitsin SV, Hooper DC (2001) Prospects for the treatment of multiple sclerosis by raising serum levels of uric acid, a scavenger of peroxynitrite. *Ann Neurol* 49:139 (letter)
  6. Drulovic J, Dujmovic I, Stojavljevic N et al (2001) Uric acid level in sera from patients with multiple sclerosis. *J Neurol* 248:121–126
  7. Poser CM, Paty DW, Scheinberg L et al (1983) New diagnostic criteria for multiple sclerosis: guidelines for research protocols. *Ann Neurol* 13:227–231
  8. Langford HG, Blaurock MD, Borhani NO et al (1987) Is thi-azide-produced uric acid elevation harmful? *Arch Intern Med* 147:645–649
  9. Jelic-Ivanovic Z, Spasic S, Majkic-Singh N, Todorovic P (1985) Effects of some anti-inflammatory drugs on 12 blood constituents: protocol for the study of in vivo effects of drugs. *Clin Chem* 31:1141–1143
  10. Ring HA, Heller AJ, Marshall WJ, Johnson AL, Reynolds EH (1991) Plasma uric acid in patients receiving anticonvulsant monotherapy. *Epilepsy Res* 8:241–244
  11. Van der Veen RC, Hinton DR, Incardonna F, Hofman FM (1997) Extensive peroxynitrite activity during progressive stages of central nervous system inflammation. *J Neuroimmunol* 77:1–7
  12. Radi R, Beckman JS, Bush KM, Freeman BA (1991) Peroxynitrite-induced membrane lipid peroxidation: the cytotoxic potential of superoxide and nitric oxide. *Arch Biochem Biophys* 288:481–487
  13. Vladimirova O, O'Connor J, Cahill A, Alder H, Butunoi C, Kalman B (1998) Oxidative damage to DNA in plaques of MS brains. *Mult Scler* 4:413–418
  14. Whiteman M, Halliwell B (1996) Protection against peroxynitrite-dependent tyrosine nitration and alpha 1-proteinase inactivation by ascorbic acid. A comparison with other antioxidants. *Free Radical Res* 25:275–283.
  15. Ischiropoulos H, Zhu L, Chen J et al (1992) Peroxynitrite-mediated tyrosinase nitration catalyzed by superoxide dismutase. *Arch Biochem Biophys* 298:431–437
  16. Hooper DC, Ohnishi ST, Kean R, Numagami Y, Dietzschold B, Koprowski H (1995) Local nitric oxide production in viral and autoimmune diseases of the central nervous system. *Proc Natl Acad Sci USA* 92:5312–5316
  17. Boullerne AI, Petry KG, Meynard M, Geffard M (1995) Indirect evidence for nitric oxide involvement in multiple sclerosis by characterization of circulating antibodies directed against conjugated S-nitrocyteine. *J Neuroimmunol* 60:117–124
  18. Newcombe J, Li H, Curzner ML (1994) Low density lipoprotein uptake by macrophages in multiple sclerosis plaques: implication for pathogenesis. *Neuropathol Appl Neurobiol* 20:152–162
  19. LeVine SM, Wetzel DL (1998) Chemical analysis of multiple sclerosis lesions by FT-IR microspectroscopy. *Free Radic Biol Med* 25:33–41
  20. Touil T, Deloire-Grassin MS, Vital C, Petry KG, Brochet B (2001) In vivo damage of CNS myelin and axons induced by peroxynitrite. *Neuroreport* 12:3637–3644
  21. Brundin L, Morcos E, Olsson T, Wiklund NP, Andersson M (1999) Increased intrathecal nitric oxide formation in multiple sclerosis; cerebrospinal fluid nitrite as activity marker. *Eur J Neurol* 6:585–590
  22. Yamashita T, Ando Y, Obayashi K, Uchino M, Ando M (1997) Changes in nitrite and nitrate (NO<sub>2</sub>/NO<sub>3</sub><sup>-</sup>) levels in cerebrospinal fluid of patients with multiple sclerosis. *J Neurol Sci* 153:32–34
  23. de Bustos F, Navarro JA, de Andres C et al (1999) Cerebrospinal fluid nitrate levels in patients with multiple sclerosis. *Eur Neurol* 41:44–47
  24. Giovannoni G, Silver NC, O'Riordan J et al (1999) Increased urinary nitric oxide metabolites in patients with multiple sclerosis correlates with early and relapsing disease. *Mult Scler* 5:335–341
  25. Santiago E, Perez-Mediavilla LA, Lopez-Moratalla N (1998) The role of nitric oxide in the pathogenesis of multiple sclerosis. *J Physiol Biochem* 54:229–237
  26. Kieseier BC, Storch MK, Archelos JJ, Martino G, Hartung HP (1999) Effector pathways in immune mediated central nervous system demyelination. *Curr Opin Neurol* 12:323–336
  27. Koprowski H, Zheng YM, Heber-Katz E et al (1993) In vivo expression of inducible nitric oxide synthase in experimentally induced neurologic disease. *Proc Natl Acad Sci USA* 90:3024–3027
  28. Schmidt HW, Walter U (1994) NO at work. *Cell* 78:919–925
  29. Xu L, Huang Y, Yang J et al (1999) Dendritic cell-derived nitric oxide is involved in IL-4-induced suppression of experimental allergic encephalomyelitis (EAE) in lewis rats. *Clin Exp Immunol* 118:115–121
  30. Lorsbach RB, Murphy WJ, Lowenstein CJ, Snyder SH, Russell SW (1993) Expression of the nitric-oxide synthase gene in mouse macrophages activated for tumor cell killing. Molecular basis for the synergy between interferon-γ and lipopolysaccharide. *J Biol Chem* 268:1908–1913
  31. Hua LL, Liu JS, Brosnan CF, Lee SC (1998) Selective inhibition of human glial inducible nitric oxide synthase by interferon-beta: implication for multiple sclerosis. *Ann Neurol* 43:384–387
  32. Cross AH, Misko TP, Lin RF, Hickey WF, Trotter JL, Tilton RG (1994) Aminoguanidine, an inhibitor of inducible nitric oxide synthase, ameliorates experimental autoimmune encephalomyelitis in SJL mice. *J Clin Invest* 93:2684–2690
  33. Stanislaus R, Pahan K, Singh AK, Singh I (1999) Amelioration of experimental allergic encephalomyelitis in lewis rats by lovastatin. *Neurosci Lett* 269:71–74
  34. Ding M, Zhang M, Wong JL, Rogers NE, Ignarro LJ, Voskuhl RR (1998) Antisense knockdown of inducible nitric oxide synthase inhibits induction of experimental autoimmune encephalomyelitis in SJL/J mice. *J Immunol* 160:2560–2564
  35. Fenyk-Melody JE, Garrison AE, Brunnert SR et al (1998) Experimental autoimmune encephalomyelitis is exacerbated in mice lacking the NOS2 gene. *J Immunol* 160:2940–2946
  36. Benveniste EN (1997) Role of macrophages/microglia in multiple sclerosis and experimental allergic encephalomyelitis. *J Mol Med* 75:165–173
  37. Marchetti B, Morale MC, Brouwer J et al (2002) Exposure to a dysfunctional glucocorticoid receptor from early embryonic life programs the resistance to experimental autoimmune encephalomyelitis via nitric oxide-induced immunosuppression. *J Immunol* 168:5848–5859

38. Xu C, Hillert J (1998) Absence of linkage with the neuronal nitric oxide synthase (NOS1) gene in 41 multiplex Swedish MS families. *Eur J Neurol* 5:393–396
39. Honegger CG, Krenger W, Langemann H (1989) Measurement of free radical scavengers in the spinal cord of rats with experimental autoimmune encephalomyelitis. *Neurosci Lett* 18:327–332.
40. Langemann H, Kabiersh A, Newcombe J (1992) Measurement of low-molecular-weight antioxidant, uric acid, tyrosine and tryptophan in plaques and white matter from patients with multiple sclerosis. *Eur Neurol* 32:248–252
41. Spitsin S, Hooper DC, Mikheeva T, Koprowski H (2001) Uric acid levels in patients with multiple sclerosis: analysis in mono- and dizygotic twins. *Mult Scler* 7:165–166
42. Costantinescu CS, Freitag P, Kappos L (2000) Increase in serum levels of uric acid, an endogenous antioxidant, under treatment with glatiramer acetate for multiple sclerosis. *Mult Scler* 6:378–381
43. Kock R, Delvoux B, Sigmund M, Greiling H (1994) A comparative study of the concentrations of hypoxanthine, xanthine, uric acid and allantoin in the peripheral blood of normals and patients with acute myocardial infarction and other ischaemic diseases. *Eur J Clin Chem Clin Biochem* 32:837–842
44. Ogiwara T, Kim HS, Hirano K et al (1998) Oxidation products of uric acid and ascorbic acid in preterm infants with chronic lung disease. *Biol Neonate* 73:24–33
45. Hellsten Y, Tullson PC, Richter EA, Bangsbo J (1997) Oxidation of urate in human skeletal muscle during exercise. *Free Radic Biol Med* 22:169–174
46. Spitsin S, Hooper DC, Leist T, Streletz LJ, Mikheeva T, Koprowski H (2001) Inactivation of peroxynitrite in multiple sclerosis patients after oral administration of inosine may suggest possible approaches to therapy of the disease. *Mult Scler* 7:313–319
47. Toncev G, Milicic B, Toncev S, Samardzic G (2002) Serum uric acid levels in multiple sclerosis patients correlate with activity of disease and blood-brain barrier dysfunction. *Eur J Neurol* 9:221–226