

Vitamin D

Second Edition

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CHAPTER 61

The Pharmacology of Vitamin D, Including Fortification Strategies

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INTRODUCTION

The use of vitamin D has never been approached in the same way that we would expect to see for any modern drug. Unlike other nutrients, we have never had dietary intakes of vitamin D as a reasonable reference point for deciding on how much of this nutrient/drug that people should be consuming. The ambiguity between “nutrient” and “drug” is reasonable when it comes to vitamin D, because there were no meaningful amounts of vitamin D in the kinds of foods that Paleolithic humans would likely have been consuming.

Our biology was designed by evolution for life in equatorial Africa. Therefore, consumption of those rare foods that do contain a meaningful amount of vitamin D, like ocean fish, could not have played a role in determining human vitamin D requirements. Requirements for vitamin D were satisfied by the life of the naked ape that became the species, *homo sapien*, in its native, tropical environment. Since our culture and environment no longer match the conditions that defined our biology, we modern humans might benefit if we could compensate for the biological consequences of modern life. One such consequence may be an endemic lack of vitamin D that can be corrected by appropriate supplementation.

My perspective is the North American one, where vitamin D is primarily regarded as a nutrient. However, in Europe and the rest of the world, use of even small doses of vitamin D usually falls into the realm of a prescription drug. That perspective has the advantage of imposing a higher expectation on our understanding of the use of vitamin D. Before approving the clinical use of any new drug, government regulators expect to see the answers to some relatively standard questions. Pharmaceutical firms need to anticipate these issues as they plan the research necessary for implementation of

new products. These questions include, but are not limited to, the following:

- 1a. What is the disease indication for the drug?
- 1b. What kind of clinical or health effects should we be looking for, based on preclinical animal and laboratory research?
- 2a. What are the most useful approaches to delivering the drug to people: the vehicle?
- 2b. What is the appropriate dosage, route of administration, and interval between doses?
3. What is the desirable target for the plasma concentration, what dose would be needed to attain or ensure this?
4. What, if any, are the biological markers to monitor toxicity, and what are our criteria for determining therapeutic effectiveness? What is the "therapeutic index" the ratio between toxic vs beneficial dose levels?

When it comes to plain and simple, nutritional vitamin D, the answer to each of these questions is that we have just started to address it in the past decade. Any opinion about vitamin D here is controversial. In an effort to provide some answers to the preceding questions, I will present perspectives about the vitamin D system that relate to pharmacological aspects of vitamin D in the adult context.

FRACTURE-PREVENTION STUDIES WITH VITAMIN D3

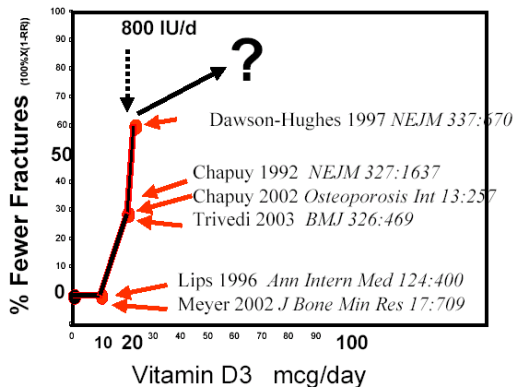


Figure 1. Summary of randomized-control clinical trials of fracture-prevention using vitamin D, with or without calcium. None of the studies using doses of vitamin D3 providing less than 20 mcg/day was effective in reducing fracture risk (80;124;124). However, all the studies in which there was a reduction in fracture risk used approximately 20 mcg/day of vitamin D3 (2;5-7)(122;122;124). This dose includes the background intake; for the work by Dawson-Hughes background intake, this was 5 mcg/day (6).

INDICATIONS AND CLINICAL USE: POTENTIAL HEALTH EFFECTS OF VITAMIN D

The only officially recognized indications for use of vitamin D in the adult are the prevention of bone loss and fractures. **Figure 1** summarizes randomized control trials looking at whether vitamin D, with or without calcium affects risk of non-vertebral fracture. A recent, thorough review of vitamin D, 1,25(OH)₂D and its analogs is also available, addressing the issue of osteoporosis prevention and treatment (1). The purpose of Figure 1 is to show that there has been no evidence that doses of vitamin D less than 800 IU/day are effective in preventing osteoporotic fractures.

Whether or not additional calcium is needed in concert with vitamin D is difficult to tell, because most studies have combined calcium and vitamin D for comparison to a placebo group receiving neither. There are now two randomized-controlled studies show that vitamin D₃ given by itself in doses of 100,000 IU (2500 mcg) every 4 months (2), or annually (3) reduces the occurrence of fractures.

Bone density declines more quickly during winter than during summer. Vitamin D supplements (about 20 mcg (800 IU) per day) combined with calcium, eliminate the faster fall in bone density during winter (4). Furthermore, three studies show that the combination of calcium and 20 mcg vitamin D together lower fracture risk in adults older than age 65 (5-7).

Occurrence of fractures is reduced by about a third, even within the first year of these studies, when bone density is not increased by enough to account for the fewer fractures (6). The explanation for this may be that vitamin D improves muscle strength and balance. This reduces the occurrence of the falls that cause fractures (8-10).

In people younger than age 65, risk of osteoporotic fracture is difficult to assess because it is a rare event prior to age 70. Data from the Nurses Health

Study suggest a 37% lower risk of osteoporotic fracture in postmenopausal women younger than 65, if they consume vitamin D in amounts of at least 12.5 mcg/day, compared to women consuming less than 3.5 mcg/day vitamin D (11). The authors failed detect any effect of calcium intake, but suggested that in this cross-sectional study, women with a family history of osteoporosis would have been more likely to take calcium, confounding a calcium effect.

I have a concern that 1,25(OH)₂D may be used too aggressively as an alternative to improved vitamin D nutrition, in the prevention or treatment of osteoporosis. The point that 1,25(OH)₂D has a narrower margin of safety (therapeutic index) than vitamin D has never been raised in analyses comparing them (1;12). If osteoporosis occurs because the vitamin D system is somehow deficient or defective, it makes little sense to resort to the use of 1,25(OH)₂D. Rickets and osteomalacia usually exist despite normal 1,25(OH)₂D concentrations. Increases in vitamin D will not increase 1,25(OH)₂D levels further (13-16). As kidney function deteriorates, its endocrine capability also declines, and thus a low serum 1,25(OH)₂D level reflects impaired renal function, not poor nutrition (16;17). Whatever effect that aging *per se* has on 1,25(OH)₂D levels, this can be overcome by increasing the 25(OH)D concentration (18). Despite many studies looking into the use of 1,25(OH)₂D and its analogs to prevent or treat osteoporosis, the review of this topic by Papadimitropoulos concludes that there no reason for anyone to resort to any metabolite other than nutritional vitamin D (1). I would add that this must be vitamin D₃, and at a dose of at least 20 mcg/d.

Non-Bone Effects Of Vitamin D

Vitamin D nutrition probably affects health beyond just bone. The mechanisms involved in mediating the non-classic (i.e. non-bone) effects of vitamin D are probably through 1,25(OH)₂D produced locally, using circulating 25(OH)D as the substrate. Many tissues possess 25(OH)D-1-alpha-hydroxylase, including the skin (basal keratinocytes, and hair follicles), lymph nodes (granulomata), pancreas (islets), adrenal medulla, brain, pancreas, and colon (19). An even wider range of tissues possess receptors for 1,25(OH)₂D (VDR) (20). All of this reveals a system for paracrine regulation of tissue processes that involves the local production of 1,25(OH)₂D. Sufficient vitamin D nutrition, and hence, appropriate 25(OH)D concentration is essential to this local, paracrine role of 1,25(OH)₂D that is not generally reflected in the circulating level of 1,25(OH)₂D. The paracrine components of the vitamin D endocrine/paracrine systems account for the many effects of vitamin D nutrition and/or UVB light on health and disease prevention.

While all of the effects in Table 1 are statistically significant, most of the evidence for a role of vitamin D is circumstantial. Epidemiological studies show that higher serum 25(OH)D, and/or environmental ultraviolet exposure is associated with lower rates of breast, ovarian, prostate, and colorectal cancers (21-28). More recent statistical analyses also show significant relationships including non-Hodgkin's lymphoma, and cancer of the bladder, esophagus, kidney, lung, pancreas, rectum, stomach and corpus uteri (29). Multiple sclerosis is more prevalent in populations having lower levels of vitamin D nutrition or ultraviolet exposure (26;30-32), and it has been proposed that vitamin D intake, ranging from

1,300 to 3,800 units per day, helps prevent the disease (32). Established osteoarthritis progresses more slowly (is less severe) in adults with higher vitamin D nutritional status, with serum 25(OH)D that exceeds 75 nmol/L (33;34). The prevalence of hypertension increases with population distance, north or south, from the equator (35). Blood pressure is lowered in mildly hypertensive patients whose 25(OH)D levels are raised to over 100 nmol/L by tanning (36). One randomized intervention study showing that vitamin D supplementation at 20 µg/d (800 IU/d) lowers blood pressure in elderly women (37). Vitamin D deficiency impairs immune function in animals (38), and in children there is a strong association between pneumonia and nutritional rickets (39). The concept that there is a connection between vitamin D nutrition and immune function is further supported by the apparent protective effect of improved vitamin D nutrition during infancy and childhood against type I diabetes mellitus (40). If any of these non-traditional effects of vitamin D were taken into account, they would result in a substantial upward revision of the AI (RDA) for vitamin D.

The level of evidence needed to make a health claim that can be sanctioned officially involves more than the circumstantial evidence of laboratory experiments and epidemiology. It requires direct intervention, the controlled administration of the agent to many healthy people, and showing an effect that stands up to statistical analysis. We need randomized intervention trials to take this field beyond pre-clinical basic research and epidemiological evidence. There are ongoing randomized trials involving “vitamin D” that relate to cancer, multiple sclerosis, and osteoporosis, but for the most part, they deal with analogs of 1,25(OH)2D, not the nutrient.

The nutrient has been very much overlooked for all purposes except rickets, osteomalacia and osteoporosis. There are three reasons for this. First, the financial incentive lies with the proprietary analogs, driven by private funding that diverts the focus of investigators who are able to do such studies. Second, an optimized dose of vitamin D has never been established for adults. Therefore, “plain” vitamin D sometimes compares poorly with 1,25(OH)2D and its analogs whose dose is more thoroughly optimized (12), and whose dose is usually designed to be very close to the point where it could cause hypercalcemia. As I will discuss later, optimal doses of vitamin D probably vary, depending on the indication, so that one dose may not always be optimal. Third, the official mis-representation that vitamin D2 and vitamin D3 are equal has resulted in efficacy studies at higher doses usually involving vitamin D2 because high-dose commercial preparations of vitamin D are comprised of this. One example of this is work looking at whether vitamin D2 supplementation might prevent bone loss in steroid-treated patients (41;42); the effects of “vitamin D” were marginal, but since plain and simple vitamin D3 was never part of the experimental protocol, the issue remains unresolved. Another example of the unfortunate focus on vitamin D2 instead of the D3 form is the recent Australian study using vitamin D2 at a decent dose, 10,000 IU (250 mcg) weekly, producing no significant effect on bone density preservation, but showing essentially no effect on serum 25(OH)D either (43).

OVERVIEW OF THE SYSTEM OF VITAMIN D METABOLISM, AND ITS REGULATION.

Administration of vitamin D (cholecalciferol) is unusual in pharmacology or in endocrinology, because this molecule is two metabolic steps away from the biologically active agent, 1,25(OH)2D. Furthermore, the laboratory test to monitor dose is the concentration of a metabolite, 25(OH)D. Figure 2 illustrates the metabolite “compartments” occupied by vitamin D after ingestion or exposure to sunshine. Less than the 25 percent of vitamin D that enters the body actually becomes 25(OH)D. More than 75 percent of vitamin D entering the circulation bypasses what we recognize as the vitamin D endocrine system. Instead, most vitamin D entering the circulation is excreted and/or metabolized by other routes not shown here, and most likely, excreted into the bile.

Figure 2 consists of two panels to illustrate the metabolic adaptations that exist so that the vitamin D endocrine system can accommodate to a wide range in the substrate concentration. The vitamin D system is optimized to maintain plasma 1,25(OH)2D levels according to the requirements of calcium homeostasis. The earliest compromise to progressive restriction in vitamin D supply is a diminished capacity of non-renal tissues to produce 1,25(OH)2D. This compromise at non-renal tissues is illustrated in Figure 2 by the greater height of one of the valves representing the 1-hydroxylase on the “pail” that represents the 25(OH)D compartment.

If one looks at the system of vitamin D metabolism in Figure 2 from the perspective of a system designed to control something, it becomes clear that this is a system better designed to cope with an abundance of supply, not a lack of it. The flow of vitamin D toward 25(OH)D is remarkably inefficient, with most bypassing it. Furthermore, there is no way to correct for deficiency of vitamin D, other than to redirect utilization of 25(OH)D toward 1,25(OH)2D production, which is the pathway most acutely important for life. That is, when supplies of vitamin D are severely restricted, its metabolism is directed only toward the maintenance of calcium homeostasis. To expand on the point that the system of vitamin D metabolism is effectively a designed for adjusting for higher inputs, not lower inputs, I offer the example of an air-conditioner system. Air conditioners are designed to compensate for excessive heat, but they are a useless way to compensate for a cold environment. The environment under which human vitamin D metabolism was effectively designed was for people without clothing, living at latitudes where UVB intensity was always enough to produce a relative abundance of vitamin D. In contrast, most modern humans cover close to 95 percent of skin surface and avoid sunshine. The vitamin D endocrine/paracrine system is not designed to cope with the lack of vitamin D created by our modern culture of clothing and sun-avoidance. Inadequate supplies of vitamin D limit the paracrine control that many tissues need so they can function properly. As a result, it is possible that what we regard as a modern “normal” prevalence of some of the diseases listed in Table 1 could be reduced substantially if we were to increase our intakes of vitamin D (44-47).

Control of metabolism in the vitamin D endocrine system is very different from the way other steroid hormones are regulated. For conventional steroid hormones, the concentration of substrate (cholesterol) is far higher than the substrate in the vitamin D system. Figure 3 illustrates the effective *in-vivo* Km of 1-hydroxylase, in relation to the physiological concentration

range of its substrate. Our circulating cholesterol concentration is in the order of 5 million nmol/liter; in contrast, 25(OH)D typically circulates at less than 200 nmol/liter. Cholesterol concentration is not a rate-limiting aspect of the body's capacity to generate steroid hormones; however, 25(OH)D concentration is absolutely rate-limiting for 1,25(OH)2D production.

In the acute situation, before adjustments can be made to 24-hydroxylase and catabolic pathways (before the "valves" in Figure 2 can be adjusted), the *in vivo* production of 1,25(OH)2D is directly proportional to circulating 25(OH)D concentration. In rats, the acute injection of 25(OH)D into the circulation produces a rapid increase in 1,25(OH)2D, directly proportional to the percentage increase in 25(OH)D (48;49). Since *in vivo* concentrations of 25(OH)D change slowly, over many days and weeks, this first-order relationship between 25(OH)D and 1,25(OH)2D is not normally seen in adults (18). However, in situations where 1-hydroxylase is tonically stimulated, either because of primary hyperparathyroidism (50) or in granulomatous disease (51;52), modest increases in vitamin D supply will raise plasma 1,25(OH)2D concentration and aggravate hypercalcemia.

Differences between steroid hormones and the vitamin D system are amplified further by the large differences in concentrations of their respective plasma carrier proteins. Sex-steroid binding globulin, and glucocorticoid binding globulin circulate in concentrations of about 200 nmol/L, in the same order of magnitude as their ligands (53); in contrast, the concentration of vitamin D binding protein is 4700 nmol/L (54); this represents a 50-fold excess over its vitamin D-derived ligands.

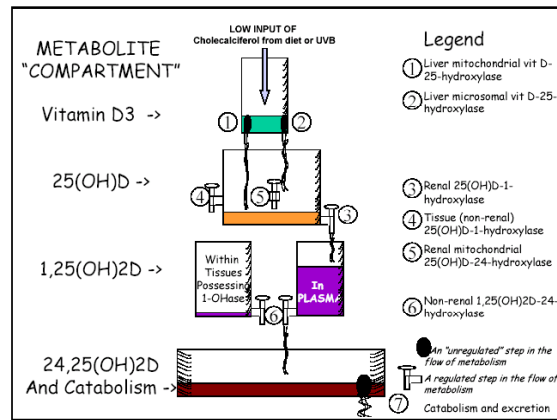


Figure 2A. Metabolism of vitamin D under conditions of low vitamin D supply. The vessels represent metabolic compartments, stages in the metabolism of vitamin D. The height of the shaded portion of each vessel represents the relative concentration of each metabolite indicated in the figure. This figure illustrates the concept that vitamin D metabolism *in vivo* functions below its K_m , i.e. the system behaves according to the first-order reaction kinetics. Just as the flow of water through a hole in a pail reflects the height of water in that pail, the rates of metabolism in the vitamin D system reflect the concentration of precursor at each step. Open passages represent steps in metabolism in which the pertinent enzymes are relatively unregulated. Valves represent steps in metabolism in which there is regulation of flow at the enzyme (this regulation is usually through changes in amount of enzyme protein in specific tissues, and not allosteric). When vitamin D supplies are low, flow of 25(OH)D through other potential pathways is compromised to maintain the circulating concentration of 1,25(OH)2D at the level determined by the priority requirements of bone and mineral metabolism.

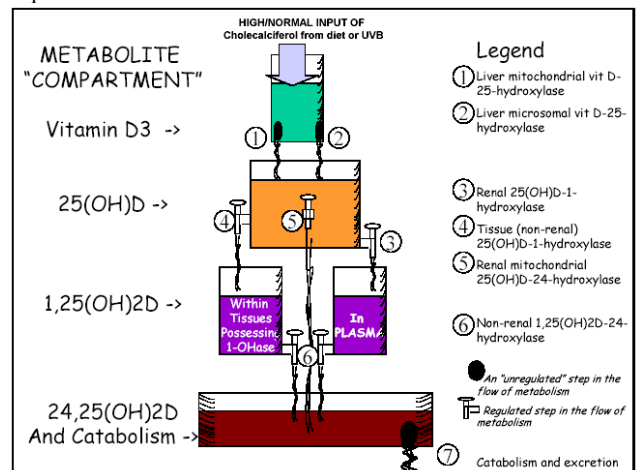


Figure 2B. Metabolism of vitamin D under conditions of adequate vitamin D supply. When vitamin D supplies are adequate, flow of 25(OH)D through other potential pathways, including its utilization by peripheral tissues for paracrine regulation, is no longer compromised. Higher 25(OH)D concentration makes available routes of metabolism other than the one path needed for bone and mineral metabolism. Furthermore, a higher supply of vitamin D leads to an upregulation of 24-hydroxylase and the catabolic pathways associated with it, this accelerates rate of metabolic clearance and metabolite turnover in each compartment.

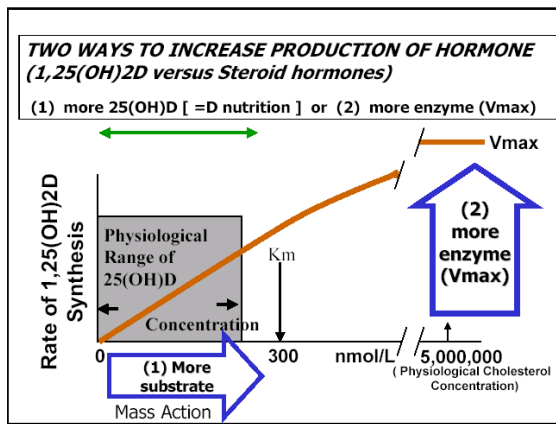


Figure 3. The difference in enzyme kinetics between the vitamin D endocrine system and the substrate supply for conventional steroid hormone systems based on cholesterol. The purpose of this figure is to emphasize that the range of physiologic concentration of 25(OH)D in mammals is less than the Michaelis-Menten constant (K_m) of 1-hydroxylase that has been characterized in vitro (125), and in vivo (48). There are two ways to improve capacity for 1,25(OH)₂D production at kidney, and at peripheral tissues: provide more substrate, or increase 1-hydroxylase content of the tissue. This is fundamentally different from the situation relevant to every other part of the endocrine system. No other hormone is so dependent on the arbitrary, external supply of its structural raw material. The concept of a mass-action relationship for 1,25(OH)₂D production is the basis of the argument that operation of paracrine control systems dependent on vitamin D supply can be improved by improving vitamin D nutrition.

The dynamics of 25(OH)D in tissues are remarkable. Its carrier protein, DBP, is cleared from plasma with a half-life of 1.7 d, which is shorter than the 5-day half life of albumin (55). Within one hour after injection of radiolabeled DBP, the radiolabel is present in a greater concentration than in plasma, within kidney, liver, skeletal muscle, heart, lung, intestine, testis, and bone (55). In contrast to DBP, its ligand, 25(OH)D, is cleared slowly from the body, with a half-life of about 10 days in both rabbit (55) and human (56). The pool of DBP outside plasma is double the size of the intravascular DBP pool, and the molar replacement rate of DBP is reported to be 1,350-fold higher than that of 25(OH)D. The binding of 25(OH)D to DBP does not affect the turnover or tissue uptake of DBP (55).

As a short summary of the preceding, the DBP and/or DBP-25(OH)D₃ complex is removed from plasma by a variety of tissues. The DBP is degraded during this process, and most 25(OH)D released within those tissues is recycled. The molar excess of DBP to 25(OH)D in plasma, and the relatively rapid turnover of DBP indicate that a high capacity, high affinity, and dynamic transport mechanism for vitamin D sterols exists in plasma. Most 25(OH)D released into cells because of the metabolic clearance of DBP is recycled; however, the clearance of DBP also provides ready access to vitamin D, 25(OH)D and its metabolites by the liver and kidney, the two organs most involved in the clearance of DBP, and which are the two organs central to the endocrine function of the vitamin D system.

Recent new knowledge of the megalin/cubulin system has shed light on the mechanisms of DBP-tissue interactions, and tissue-specific uptake of DBP (chapters 8-10 of this book) (57). Megalin and cubulin are cell-surface, endocytic receptors, members of the low-density lipoprotein receptor gene family. These help to regulate the concentration of ligands in the extracellular fluids and deliver metabolites to cells in need of these metabolites (58). Differences in tissue distribution of these cell-surface proteins will affect the accessibility of different tissues to circulating 25(OH)D.

DOSAGE CONSIDERATIONS

Infants

Cholecalciferol, or vitamin D₃, given in the form of cod liver oil, has been a folk remedy in northern Europe since the 1700's. Empirically, a small teaspoon-full daily was thought to help infants thrive. This arbitrary dose of cod-liver oil has turned out to be a good guess, so far as infants are concerned. The 375 IU (9 mcg) of vitamin D₃ contained in that teaspoon (59) was confirmed relatively recently as being appropriate for infants (60;61). A French study utilizing vitamin D₂ concluded that neonates might need somewhat more, 1000 IU (62). If safety of vitamin D during infancy is a concern, it should be kept in mind, that until the late 1960's, the recommended amount of vitamin D for infants in Finland was 2000 IU/day (50 mcg/day). A large epidemiologic study suggests that this higher dose lowered risk of juvenile diabetes before age 30 years, by 85% compared to people not receiving vitamin D as infants (40).

Compared to the adult, vitamin D nutrition in the infant and child has been well characterized, and it is the focus of Chapter 65 (Pettifor). There is also an excellent review of the field available, by Chesney (63). The present chapter focuses on the pharmacology of vitamin D in the adult.

Adults

Until it became clear that vitamin D was important to the health of adults, there was very little thought directed at how much vitamin D adults might need to consume. Until recently, there was been no consensus about what the objective criteria should be for appropriate vitamin D nutrition. In England, an adult recommendation of 2.5 mcg (100 IU)/d was set simply because 7 women with severe nutritional osteomalacia showed a response to this amount (64).

Interestingly, the oils of different fish contain different amounts of vitamin D. For example, a teaspoon full of halibut liver oil contains twice as much vitamin D₃ as does cod liver oil. If it had been halibut liver oil used in the past, recommendations for vitamin D supplementation could well have been double what they have been through most of the last century.

Into the 1960s, the absence of overt rickets or osteomalacia was the only criterion that vitamin D nutrition was adequate (65). By the same criterion, anthropologists also consider vitamin D nutrition to have been a relatively minor problem for ancient populations. This is now explained by the new concept that the lack of vitamin D resulted in a natural selection for white skin colour as a way to prevent rickets and osteomalacia within defined environments (66). Women with osteomalacia would have produced few offspring, while those able to produce enough vitamin D to prevent rickets and

osteomalacia would have been the vast majority in any region - survival depended on adequacy of vitamin D nutrition, and natural selection of skin color helped to ensure adequacy.

In the 1960s, an expert committee on vitamin D could provide only anecdotal support for “the hypothesis of a small requirement” for vitamin D in adults and recommended one-half the infant dose, to ensure that adults obtain some from the diet (65). Despite the new knowledge uncovered since that time, dietary vitamin D recommendations for adults have remained very conservative, and still derive from amounts established for neonates. In contrast to the way decisions are made about the dose of any new drug, recommendations for vitamin D have been arbitrary, because there was no firm evidence on which to base decisions. However, even though the evidence about the effects of vitamin D dosages on adult health have become characterized scientifically, those with the final say in setting official nutrient guidelines (not the experts they consulted) continued to focus on lower doses of vitamin D than had been shown effective in the fracture prevention trials discussed above. The revised recommendations were referred to as the “adequate intake” (AI), because there was no published evidence of efficacy for them (67;68).

The objective measure of vitamin D nutritional status is the 25-hydroxyvitamin D (25(OH)D) concentration in serum or plasma (67). The consensus on this point has made it possible for researchers to focus on a measurable target when it comes to vitamin D nutrition. **Table 2** summarizes two views of the relationships between long-term vitamin D intakes and the anticipated range of 25(OH)D concentration associated with them.

Figure 4 is a dose-response curve to showing the final average 25(OH)D concentrations attained in studies reported in the literature (44;69). **Table 3** summarizes incremental responses to different treatment strategies to raise 25(OH)D to steady-state concentrations. Responsiveness to vitamin D administration, as measured by the nmol per liter increase per mcg consumption per day, increases with: a) lower vitamin D dosage, b) lower initial 25(OH)D concentration; c) longer duration of supplementation, suggesting a long half-life and time to plateau.

The conventional approach to improving vitamin D nutritional status has been to give either vitamin D3 or vitamin D2 (ergocalciferol). Until recently, availability of 25(OH)D was another option (supply of this product has been discontinued by Organon, NJ, USA). The company’s discontinuation of 25(OH)D may have made sense, because the objective of increasing plasma 25(OH)D concentrations can be almost as easily achieved by providing enough vitamin D3. Nonetheless, useful perspectives can be gained from previous experience with 25(OH)D. Barger-Lux and Heaney et al. have shown that as 25(OH)D dosage increases, there is effectively a linear increase in the average 25(OH)D concentration achieved (**Table 3**). However, when vitamin D3 is used, the increment in 25(OH)D per mcg per day of vitamin D3 decreases as dose increases.



Table 1. Diseases and conditions known to be, or implicated as being prevented by greater vitamin D nutrition or skin UV exposure

<i>Disease</i>	<i>Type of evidence supporting the association</i>	Reference
Rickets	Long established, preventive	
Osteomalacia	Long established, preventive	
Osteoporosis	Placebo-controlled, randomized studies that vitamin D prevents loss of bone density, and lessens fracture risk	(2;4-6)
Poor calcium absorption	Modest increase in Vitamin D nutrition increases this.	Heaney2003dietetic j
blood-pressure regulation	Epidemiological and randomized interventional data	(8;9;35;36)
Risk of diabetes	epidemiological and case-control data	(40;126;127)
progression osteoarthritis	epidemiological, cross-sectional studies	(33;34)
diminished intra-uterine growth	Presumed effect	(128),
Effects on brain development	Rat experiments	(129)
resistance to pneumonia,	Epidemiological association with rickets	(39)
Multiple sclerosis, occurrence and progression	Epidemiological data, and lab effects on tissue	(32;130;131)
prevention of tuberculosis,	Epidemiological data, and lab effects on tissue	(132;133)
<i>Prevent Depression or SAD or Improve mood</i>	Small RCT's 400 IU/day or 100000 in winter	(134;135)
	No mood effect of 400 IU/day	(136)
Lessen risk/severity of Fibromyalgia	Cross-sectional study	(137)
<i>Protection against cancers</i>		
breast	Epidemiological data, and lab effects on tissue	(138;139)
prostate	Epidemiological, and lab effects on tissue	(140) (139;141;142)

Large bowel.	epidemiological and cross-sectional data, based on latitude and serum 25(OH)D	(138;139)
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Table 2. The clinical interpretation of serum 25(OH)D levels and the estimated intakes of vitamin D needed to ensure these levels (1 mcg = 40 IU).

	Deficiency (rickets and Osteomalacia)	Insufficiency (increased PTH secretion, osteoporosis)	Sufficiency	Desirable (suppress PTH, optimize Calcium absorption)	Toxic/Therapy (could increase urine and serum calcium)
Serum 25(OH)D nmol/L	0-25	25-40	40-100	75-160	>220
<i>Vitamin D3 mcg/day needed to reach the 25(OH)D above:</i>					
Food and nutrition board ^a	0 mcg/d	5-10	5-15	not stated	≥95
From Evidence Reviewed ^b	0-5 mcg/d	10-15	25-100	100-250	>1000 (> 40000 IU)

^a Implications drawn from current National Academy of Sciences nutritional guidelines that the stated intake will deliver the level of adequacy (67). The “adequate intake” recommendations for vitamin D vary according to age: adults <50, 5 mcg/day; 50-70 years, 10 mcg/day; > 70 Years, 15 mcg/day.

^b Based on literature (69;108;122)

TABLE 3. Strategies to increase circulating 25(OH)D concentration in adults: Effects of compound, dose, and duration¹.

Compound	25(OH)D increase per mcg/d	DOSE mcg/day	Duration on the dose wks	Absolute increase in 25(OH)D nmol/L	Reference
25(OH)D	4.1	50	4	206.4	(109)
25(OH)D	4.0	10	4	40	(109)
25(OH)D	3.8	20	4	76.1	(109)
cholecalciferol	1.5	15	52	22	(143)
cholecalciferol	1.4	20	8	27	(144)
cholecalciferol	1.1	25	8	28.6	(109)
cholecalciferol	1.1	21	20	23.4	(145)
cholecalciferol	0.8	100	52	81	(143)
cholecalciferol	0.8	25	20	19	(108)
cholecalciferol	0.7	138	20	102.7	(145)
cholecalciferol	0.6	275	20	169.8	(145)
cholecalciferol	0.6	250	8	146	(109)
cholecalciferol	0.5	100	20	51.8	(108)
cholecalciferol	0.5	1250	8	643	(109)
ergocalciferol	0.3	36	104		(43)

¹The results in this table represent recent work not included in Figure 4. These data were assembled to permit comparison of efficacy dose of different strategies for increasing 25(OH)D concentration. The results are sorted in order of decreasing response to the dose, based on the nmol/L increase in 25(OH)D per mcg/day of oral the doses used in these studies.

Since the increment in plasma 25(OH)D concentration per mcg dose is at least four times higher for 25(OH)D administration than for vitamin D3 administration, we can conclude that less than 25 percent of vitamin D molecules ever become 25(OH)D. At least three quarters of the molecules of vitamin D that enter the body are removed by some other fate.

The case against ergocalciferol, vitamin D2

Vitamin D is available in two forms for nutritional supplementation, ergocalciferol (vitamin D2) and cholecalciferol (vitamin D3). Vitamin D2, is manufactured by exposing a fat extract of yeast to UV light. Since no metabolite of vitamin D2 is normally detectable in the blood of humans or primates (70;71), I contend that this should be regarded as a drug, and not physiological compound. The present discussion focuses on vitamin D3, cholecalciferol, the form of vitamin D natural present in mammals. Vitamin D3 (from here on, vitamin D) is the more potent form of vitamin D in all primate species and in man (70;71). Comparisons between the two versions of vitamin D (71), and the meta-analysis of effects on 25(OH)D (Figure 4) indicate that vitamin D3 is about 4 times as potent as vitamin D2, i.e. 1 mcg of D3 = approximately 4 mcg of D2. Nonetheless, vitamin D2 continues to be used clinically as if it is equivalent, since official guidelines (67) and pharmacopeas respond slowly to new evidence.

The presumption of equivalence is based on 60-year old studies of rickets prevention in infants – evidence recognized as weak, even at the time (59;72). The older rat data suggesting that vitamin D2 and vitamin D3 were equivalent lose their meaning when it is noted that the rat-line tests last done over 50 years ago were bioassays to

establish "units" for the quantity of vitamin D not readily measured in any other way (73). With a bioassay yielding "units", equivalence is not the same as equivalence per milligram or per mole. Furthermore, all species tested show differences between the vitamin D2 and vitamin D3 (74;75). Despite these obvious problems about units, the very conservative approach to official statements has remained unchanged, that one international unit of vitamin D is equivalent to 25 nanograms of either vitamin D2 or vitamin D3 (67;73). In Australia, vitamin D3 has never been licensed for use, and the only nutritional form of vitamin D available there is vitamin D2. Strangely, at the time of this writing, there is no form of vitamin D3 commercially available in France either.

I have summarized the differences between vitamin D2 and vitamin D3 in Table 4. Based on the many major differences between the two, it is clear that unless there is some well-characterized reason to favour vitamin D2 (I am not aware of any), all use of vitamin D for nutritional and clinical purposes should specify cholecalciferol, vitamin D3.

Vitamin D is not a hormone

For most of the 20th century, there was no debate, that vitamin D was a nutrient. It was known as "the sunshine vitamin". Confusion arose when it was realized that the active form of the vitamin D molecule was 1,25(OH)2D (calcitriol), which is a hormone in the true sense of the word. A focus on the inadequacies of the term "vitamin", and a lack of consideration for the term, "hormone", led to the misconception that vitamin D itself might be a hormone instead of a nutrient. Officially

mandated nutritional committee reports in both North America (67) and Europe (76) now state that nutritional vitamin D may be more suitably referred to as a "hormone" instead of nutrient. However, vitamin D is no more a hormone than cholesterol is, because vitamin D is only the raw material needed for synthesis of 1,25(OH)2D. Practitioners not familiar with this ambiguity occasionally administer vitamin D inappropriately, when 1,25(OH)2D or an analog would be appropriate, or vice versa (anyone who specializes clinically in the field of vitamin D knows of examples of such unintended misuse). Ambiguity and the use of jargon in this field has had unfortunate consequences.

Promotion of vitamin D nutrition is hindered by alarmist reactions justifiably associated with administration of any hormone. Use of a hormone implies that natural homeostatic control is circumvented – taken over by the physician. However, vitamin D does not generate an endocrine signal, 1,25(OH)2D and its analogs do this. The purpose of supplementing with vitamin D is to optimize the natural functions of the endocrine/paracrine systems that require it.

UVB light on human skin as a dose of vitamin D.

In any discussion of vitamin D pharmacology or dosage, it would be a major oversight to ignore the role of sunshine, particularly its UVB component. As described elsewhere in this book (Chapter 3), the synthesis of vitamin D is self-limiting reaction, reaching an equilibrium after 25-20 min of summer UVB exposure for people with white skin, and producing no net increase in vitamin D production after that (77). Darker skin requires longer exposure but the potential yield of vitamin D is the

same. Exposure of full skin surface to UVB light, in an amount less than erythema, is equivalent to a vitamin D consumption of about 250 mcg (10 000 IU/d) (78-81). Lifeguards in the United States and in Israel, as well as farmers in the Caribbean all exhibit serum 25(OH)D concentrations greater than 100 nmol/L (82-84). Furthermore, even regular short periods in sun-tan parlors consistently raise serum 25(OH)D to beyond 80 nmol/L (85) (36;86;86-90). The highest 25(OH)D concentrations in the groups of adults acquiring vitamin D physiologically (via UV exposure) range up to 235 nmol/L (36;82), and none of these studies imply that such 25(OH)D levels have caused hypercalcemia. Since humans evolved as naked apes, whose native habitat was within 30 degrees latitude of the equator, I contend that our genome was selected under conditions of abundant vitamin D supply (69). As such, it is reasonable to think that the substantially lower levels of 25(OH)D prevalent among modern humans have been accompanied by biological compromises, such as increased PTH secretion and altered cellular metabolism. By now, these compromises may have been detrimental to the health of modern humans for so long, that we are no longer in a position to realize it.

Barger-Lux and Heaney studied the effect of sunshine on outdoor workers in the US Midwest, relating it to the vitamin D intakes needed to bring about the 25(OH)D levels observed. They concluded that the summertime supply from sunshine was approximately 2800 IU/d (70 mcg/d). Despite this supply during summer, it did not ensure sufficiency through the winter, when 25(OH)D was less than 50 nmol/liter in 3 of 26 subjects and less than 75 nmol/liter in 15 of 26 subjects.

Table 4. The case against vitamin D2, compared to vitamin D3

Vitamin D2	Vitamin D3	Ref
Not detectable in humans or primates unless administered from an artificial source	The natural metabolite generated within skin and the oils of fur	(146)
Vitamin D binding protein has lower affinity for vitamin D2 than for vitamin D3 and its metabolites		(147)
Generates metabolites for which there is no vitamin D3 equivalent		(148)
Microsomal 25-hydroxylase does not act on it	Substrate for both microsomal and mitochondrial 25-hydroxylases	(149;150)
Per mole of dose, 25(OH)D increases by less than with vitamin D3		(71)
The 25(OH)D response to vitamin D2 is less in the elderly than in younger adults	25(OH)D response to vitamin D3 is the same for young vs older adults	(144;151) (143)
All known cases of iatrogenic toxicity with vitamin D involved the vitamin D2 form	All known adult cases of toxicity with vitamin D3 have been unintentional, "industrial" accidents	(69;100) (111) (99)
Less stable in dose preparations		{trang1998 (71)

PHARMACOKINETIC PRINCIPLES, VOLUME OF DISTRIBUTION, TURNOVER AND HALF-LIFE AS IT PERTAINS TO VITAMIN D.

A complete understanding of the pharmacokinetics of the vitamin D system has eluded researchers. This is because of the technical issue of measuring the nanomolar quantities of vitamin D potentially embedded within tissues or excreted in catabolized forms. It is extremely difficult to detect or to measure vitamin D and its metabolites when they exist among great excesses of other lipids. Perhaps the most careful study into the fate of physiological amounts of cholecalciferol was reported by Lawson et al. (91;92). They exposed rats with shaved skin to ultraviolet light (UVB), and measured vitamin D and 25(OH)D in tissues at various times afterwards. Although adipose tissue concentration of vitamin D was never greater than the plasma concentration, it contained the largest exchangeable pool of vitamin D. Recovery of vitamin D₃ in adipose was less than 5 percent of the amount produced within the skin (91), and this low recovery was attributed to vitamin D excretion into the bile. Lawson et al estimated that the volume of distribution of unmetabolized vitamin D₃ was approximately four liters per kg (based on concentration decay curves from plasma and total amounts recovered from tissues). Vitamin D is not detectable in the adipose tissue of normal rats (92;93), but with administration of pharmacologic doses (94), or shaving of fur to increase yield 5-fold (91), vitamin D is detectable. Brouwer et al estimate the half-life of vitamin D in rat adipose tissue to be 96 days, which I find plausible because it compares with the functional half-life of 25(OH)D in humans (69). In contrast, Lawson et al estimated the vitamin D in adipose tissue to have a half-life of 13.8 days by (91). The more rapid half-life reported by Lawson et al was likely due to the younger age of the rats.

Since vitamin D is present in the body naturally, and it is not drug, it impossible to start with completely deprived individuals to do appropriate studies of pharmacokinetics. Furthermore the component of nutritional interest is 25(OH)D, and this is a metabolite of vitamin D₃. Studies using isotopic techniques show that in humans, molecules of 25(OH)D have a plasma half-life of about 10 days (56;95). However, a more practical measure of the half-life of 25(OH)D is reflected in the rate at which 25(OH)D concentrations decline upon the sudden elimination of sources of vitamin D (acute deprivation of ultraviolet light). Two studies show that when sailors embark upon 2-month-long missions in submarines, the 25(OH)D concentration decreases by approximately 50 percent (69;96;97). Follow-up of 25(OH)D concentrations in adults intoxicated with vitamin D₃ suggest that the functional *in vivo* half life is of the order of several months (98-100).

During summer, we accumulate vitamin D₃ and store it, so that supplies for vitamin D do not become completely depleted during the winter months. Within three days of a dose of vitamin D₃, very little of the original vitamin D is detectable in plasma of rats (101) or humans (102). Most vitamin D entering the circulation appears to be excreted unmetabolized into the bile. The highest total concentrations of vitamin D and its metabolites occur in plasma. However, since plasma

represents only 2.5% of body mass, larger pools of vitamin D₃ and 25(OH)D exist in adipose and muscle (103-105).

Vitamin D leaves tissue stores, and is utilized to sustain 25(OH)D concentrations over several months. When there is a continuous supply of vitamin D, like the situation for people who regularly expose a large proportion of their skin surface to tropical sunshine, an equilibrium is reached that maintains a balance between vitamin D stored within body compartments and the removal from tissue stores, its metabolism, and clearance. Under these physiologic circumstances 25(OH)D concentrations in plasma sustain levels of more than 200 nmol/L (69). At these levels of vitamin D nutrition, there has never been a concern raised that sudden loss of adipose tissue would either raise 25(OH)D or predispose to vitamin D toxicity. Likewise, despite 70 years of experience with the use of super-physiological amounts of vitamin D, there has never been a report of a sudden excess of vitamin D caused by release from adipose stores.

Storage Of Vitamin D In The Body

It is thought that since vitamin D is a fat-soluble vitamin, it must show preferential accumulation in adipose tissue (106;107). Two studies showed that following a defined dose of vitamin D or sunshine the rise in 25(OH)D was less for obese individuals than for people who weighed less. These studies did not show that adipose tissue concentrated vitamin D, and they failed to account for the obvious effect of a larger body compartment size, which should produce a lower concentration of anything, regardless of whether adipose plays a role or not. In our study using vitamin D₃ doses of 100 µg/day in adults, we found no correlation between weight and serum 25(OH)D (108). At physiological doses, cholecalciferol (unmetabolized vitamin D₃) distributes widely into tissues, not just to adipose, but to skeletal muscle and organs as well (91;105). As stated above, turnover of vitamin D stored in tissues produces a long half-life of vitamin D in the body, of about 2 months.

The amounts of vitamin D recoverable from tissue stores account for only a fraction of the dose administered (91). The animal data indicate that more than 3/4ths of the molecules of vitamin D that enter the body are catabolized and excreted without ever being stored in tissues, and without ever becoming 25(OH)D. The human data also support this. In humans, when vitamin D or 25(OH)D are given over the long-term, to achieve an equilibrium concentration of 25(OH)D, it takes more than 4 times as much vitamin D to produce the same 25(OH)D plateau (109). By definition, at that plateau in 25(OH)D, exchange of vitamin D with tissues is at equilibrium where release of stored vitamin D equals storage of new vitamin D. Still the 4-fold difference in efficacy at sustaining 25(OH)D exists when comparing effects of doses of 25(OH)D and vitamin D. That is, the difference in efficacy between 25(OH)D and vitamin D at sustaining plasma 25(OH)D concentrations cannot be explained by deposition of vitamin D into storage sites. The difference in efficacy at sustaining 25(OH)D can only explained by loss of vitamin D entering the circulation to fates other than 25-hydroxylation or storage.

VITAMIN D TOXICITY AND SAFETY ISSUES.

Amounts of vitamin D substantially greater than physiologic amounts (>250 mcg/day) are toxic because they preoccupy circulating vitamin D binding protein (DBP) and force the percent of vitamin D that is free and unbound to increase (69;110). At toxic doses, the freely circulating vitamin D, along with its metabolites, can accumulate not only in adipose (94) but also in muscle (105). The 100 µg/d vitamin D we have used in adults is physiologic and far below what would be needed to change the free fraction of vitamin D or its circulating metabolites (54). The average capacity of human plasma DBP to bind vitamin D and its metabolites is 4700 nmol/L (54), and this exceeds by 20 times the physiologic total concentration of its vitamin D-derived ligands.

The vast majority of cases of vitamin D intoxication have involved vitamin D₂ (69). The situations involving vitamin D₃, to date, have been industrial accidents (99;110;111) or poisonings from an unknown source (100). In our case, we assayed blood levels of vitamin D and its metabolites by chromatography and found that despite record-high 25(OH)D concentrations in humans (2400 nmol/liter), they were still small in comparison to a large excess of vitamin D₃ (17,000 nmol/liter) suggesting that the capacity of liver to hydroxylate vitamin D is limited.

Like anything that has an effect on living things, vitamin D can be harmful if taken in excess. I contend that the ratio of the physiologically effective dose vs the toxic level for vitamin D is similar to the safety margin of many other nutrients (including even water). The reason vitamin D has been perceived as toxic was probably because daily ingestion in the milligram range has caused harm. In contrast, milligram amounts of other nutrients are benign. Toxicity in normal adults requires intake of more than 1000 mcg/day (40,000 IU/day), which reflects amounts of vitamin D that are four times more than the 250 mcg/day can be acquired naturally by sunshine (69). In what I see as an overreaction to the potential for toxicity with vitamin D, the current recommendation (called an "Adequate Intake" in North America) for adults under age 50 represents a homeopathic dose of about 2% of what adults with white skin would be making within 20 min of summer sun. In other words, the fear of vast excess has resulted in physiologically miniscule intake recommendations for adults.

Concentrations of 1,25(OH)₂D are not increased much by vitamin D intoxication. This reflects the high level of regulation of this hormone via both its synthesis and catabolism. Nonetheless, vitamin D toxicity is probably manifest by the excessive levels of "free" 1,25(OH)₂D, displaced from its carrier protein, DBP, by the vast excess of other vitamin D metabolites (112). This excess was confirmed by studies looking into "free" 1,25(OH)₂D concentrations in vitamin D intoxicated individuals (110). This excess of metabolite over binding capacity was also confirmed by the high total of vitamin D and 25(OH)D concentrations (19500 nmol/L) in a patient intoxicated after consuming over a million units (>25 mg) daily for many months (100).

We recently reported a safety evaluation of vitamin D₃ supplementation of normal adults, involving daily consumption of 100 mcg (4,000 IU). Contrary to the report by the Narang (113) that was used by the Food and Nutrition Board to establish the 50 mcg/d (2,000 IU/day)

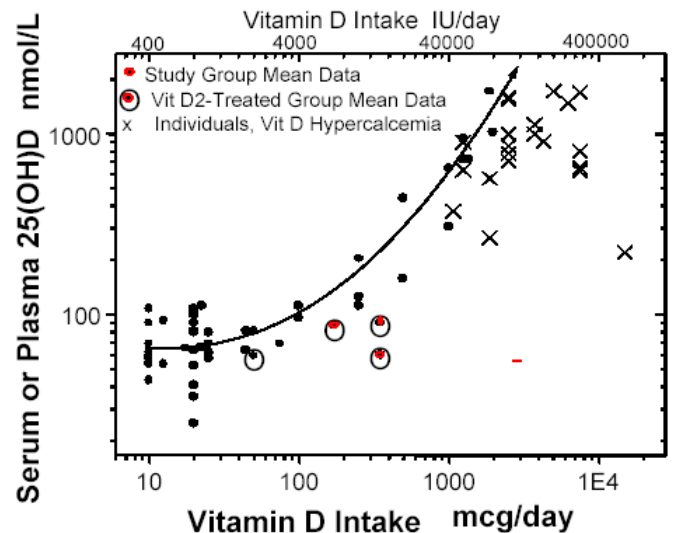


Figure 4. Dose-response relationship between daily vitamin D intake and mean 25(OH)D concentration, based on data published in the literature. The solid points show mean results for groups of adults consuming the indicated doses of vitamin D. Results for groups of adults that unambiguously consuming vitamin D₂ are shown by the circled points. Vitamin D₃ is about 4 times as potent as vitamin D₂, based on tracing the circled points for subjects consuming vitamin D₂ back to the trend-line based on vitamin D₃. Both axes are log scale. The results represented by X's are for individuals showing the classic hypercalcemic response to toxic levels of prolonged vitamin D consumption. The data used to generate this graph were compiled and published previously (44;69).

upper limit for vitamin D intake, this dose produced no detectable change in serum or urine calcium (108;114). The 25(OH)D results for this sort of study should be looked at in the context of the lowest and the highest 25(OH)D level attained with each dose. The objectives for establishing nutritional guidelines focus on the lowest level of 25(OH)D "ensured" by the given dose, while avoiding the possibility of risking an excess (108;115).

The official safety limit for vitamin D intake without supervision by a physician is referred to as the "upper limit" (UL) (114;116). This is the amount of vitamin D that the general public can take safely on a long-term basis with no anticipation of harm. Guidelines in both North America (67) and Europe (76) have established the UL as 50 µg (2000 IU) /day. This is a very conservative value that seems to remain the same, even though the evidence shows that higher intakes are safe. The value of 50 µg (2000 IU) /day has remained unchanged since it was mentioned in the 1968 Recommended Dietary Allowance publication as a dose approaching harmful level (117). To sustain the very conservative approach of making minimal changes to past recommendations, the only thing to change has been the safety margin applied, not the UL. For example when the "no observed adverse effect level" (the highest dose shown to have no harmful effect) was 2400 IU/day, based on a 1984 study (113), the safety factor applied by the United States food and nutrition Board was 1.2. When

subsequent data were published indicating that 4000 international unit/day was safe, the safety margin was increased by The European Commission to a value of 2.0 (76). Recent evidence in men shows that eight weeks of supplementation with 275 mcg/day of vitamin D does not affect circulating calcium concentration (i.e., the dose is non-calcemic, and causes no harm) (118). Even with the application of a safety factor of 2.75, this would suggest that 100 mcg (4000 IU) of vitamin D could be a safe adult UL for vitamin D. I predict that past patterns will hold true for official guidelines, and that the UL for vitamin D will remain unchanged at 50 µg (2000 IU) /day, and as a response to changing data, only the safety factor deriving that UL will change, to 5.5.

The weight of published evidence on toxicity shows that the lowest dose of vitamin D proven to cause hypercalcemia in some healthy adults is 1000 mcg (40,000 IU) per day of the vitamin D2 form (69) (Figure 4). This translates to 1000 micrograms, or 1 milligram, taken daily for many months. If a consumer wanted to achieve this toxic dose, he or she would need to take forty of the 1000 unit pills (the highest dose available in North America without a prescription) every day for many months.

Ten years ago, a dairy in the Boston area, servicing 10,000 households, made prolonged, gross errors in fortifying milk with hundreds of thousands of units (several milligrams) per quart. The case was published quickly (119) and covered by the media. The more rigorous epidemiological follow-up was published later. That showed that the situation contributed to two deaths of susceptible elderly (111). While hypercalcemia did occur, it was not widespread. By far the most susceptible group to the excess vitamin D was women over age 65 years of age, suggesting that diminished renal function may play a role. The average 25(OH)D concentration of the confirmed cases of vitamin D toxicity was 900 nmol/L (214 ng/mL) (111); in comparison, physiologically attained 25(OH)D concentrations reach 235 nmol/L safely, without hypercalcemia.

When physiologically higher vitamin D nutrition is associated with hypercalcemia, this reflects aberrant control of 25(OH)D-1-hydroxylase. This would reflect either primary hyperparathyroidism, where PTH continuously stimulates the enzyme in the kidney (50), or granulomatous disease, where peripheral tissue loses ability to regulate the 1-hydroxylase that normally serves a paracrine role (69;120).

If people with abundant sun exposure (25(OH)D > 150 nmol/L) the pre-supplement supply of vitamin D could be equivalent to about 100 mcg/d (121). An additional physiological dosage by mouth of 100 mcg/day of vitamin D would still be less than the dose of vitamin D shown to be safe in a recent study (122). As a further example of this point, we reported that 25 mcg/d of vitamin D resulted in average 25(OH)D concentrations of 69 nmol/L, while four times that amount increased 25(OH)D concentrations by only another 27 nmol/L (108). The increment with each additional amount of vitamin D becomes progressively smaller as the pre-dose 25(OH)D level increases. Thus, a further 100 mcg/d would add marginally to what I regard as the inconsequential risk due to the 250 mcg/d vitamin D supply that is physiological because it is obtainable through sun exposure. Since a long-term vitamin D consumption of at least 1000 mcg/d would be needed to

cause hypercalcemia, there is a large margin of safety with 100 mcg/d. (I would welcome any of discussion of evidence implicating harm with vitamin D3 (not D2) in adults at doses below 1000 mcg/d. There is simply nothing published about this, except in infants.)

One concern sometimes expressed, is that if adipose tissue were to break down, a sudden influx of vitamin D from adipose might be toxic (123). I want to address this issue. In both rats and cattle, high doses of vitamin D are needed before vitamin D ends up as detectable in adipose tissue (94;105). Despite being present in “significant” amounts in tissues, storage in tissues is not efficient. As a proportion of what enters the body via the skin or the diet, the amounts of vitamin D stored in adipose are a fraction of the total. In normal humans, adipose tissue content of vitamin D has been reported to be as high as 116 ng/g (approx 5 IU/g adipose) (92). In cattle intoxicated with 7.5 million IU vitamin D (to cause hypercalcemia, in an experimental process to activate proteases to make beef more tender after slaughter), muscle levels of vitamin D reached 91 ng/g tissue (4 IU/g). The highest tissue level reported in those animals was in the liver, which contained vitamin D at 979 ng/g (39 IU/g) (105). The point here is, that while there is “meaningful” storage of vitamin D in tissues, all the evidence to date indicates that only a fraction of any vitamin D dose ends up in tissues to be withdrawn at later times.

If there were a sudden breakdown of 1 Kg of adipose tissue, or liver, this would release as much as 979 mcg (39000) IU of vitamin D into the body. A toxic excess of vitamin D would require the break down daily of one Kg of adipose tissue that had been primed with by prior vitamin D intoxication, with daily adipose catabolism to continue for several weeks. When toxic doses of vitamin D are administered, the effect will be manifest during the period of administration. There is no evidence that enough residual vitamin D can be stored in adipose tissue that vitamin D toxicity could possibly arise at some later time, because of weight loss.

SUMMARY

To conclude, I return to the pharmacological questions posed at the start of this chapter, and offer Table 5 as a way to address the issues, based on the material in this chapter.

TABLE 5. OPINIONS AND BEST GUESSES AT THE ANSWERS TO PHARMACOLOGIC QUESTIONS THAT NEED TO BE ADDRESSED IN RELATION TO VITAMIN D NUTRITION

<i>Question</i>	<i>Answer</i>	
1a. What is the disease indication for the drug?	Fracture prevention; preservation of bone mineral density; normalization of PTH levels	
1b. What kind of clinical or health effects should we be looking for, based on preclinical animal and laboratory research?	Disease prevention: cancer, diabetes, multiple sclerosis, high blood pressure, fibromyalgia.	
2a. What are the most useful approaches to delivering the drug to people: the vehicle,	Encouragement to expose a large percent of skin to summertime sunshine, 10 min daily. Fortification of foods to physiologically meaningful levels of vitamin D, consumption of supplement preparations with vitamin D.	
2b. The dosage, route of administration, and	Dosage depends on target 25(OH)D desired. We can assume a rule of thumb, that vitamin D at 1 mcg/day increases 25(OH)D by 1nmol/L, after 8 months of use. Oral vitamin D is probably more effective at increasing than injection.	
2c. Interval between doses?	Since the half-life for decline in 25(OH)D is effectively 2 months, doses of vitamin D could be given monthly (we use weekly in our studies). Less frequent dosing than once every 2 months will generate large fluctuations in 25(OH)D concentrations that may not be desirable, because the enzymes involved in the regulation of vitamin D metabolism are functioning in a 1 st order relationship with substrate.	
3. What is the desirable target for the plasma concentration, what dose would be needed to attain or ensure this?	Current consensus points to a goal of ensuring that 25(OH)D levels be higher than 70-100 nmol/L. Note that to ensure this level for those with the weakest response to vitamin D, we need to aim for an average 25(OH)D concentration of about 120 nmol/L. These objectives are achieved with a dose that averages 100 mcg/day for all adults.	
4. What, if any, are the biological markers to monitor toxicity, and what are our criteria for determining therapeutic effectiveness? What is the "therapeutic index" the ratio between toxic vs beneficial dose levels?	Hypercalcemia is the classic criterion for toxicity of vitamin D and its metabolites or their analogues. "Non-calcemic" doses are considered "safe". The most sensitive clinical index of excessive vitamin D-related effects is probably hypercalciuria, which would logically occur at lower doses than hypercalcemia. Of greater concern for the long-term use of vitamin D, its metabolites or analogs should be effects on soft-tissue calcification, within aorta, kidney or other tissues. These effects may be seen radiologically in humans, or by direct measure in pre-clinical animal studies.	

Reference List

1. Papadimitropoulos E, Wells G, Shea B et al. Meta-analyses of therapies for postmenopausal osteoporosis. VIII: Meta-analysis of the efficacy of vitamin D treatment in preventing osteoporosis in postmenopausal women. *Endocr Rev* 2002; 23(4):560-569.
2. Trivedi DP, Doll R, Khaw KT. **Effect of four monthly oral vitamin D3 (cholecalciferol) supplementation on fractures and mortality in men and women living in the community: randomised double blind controlled trial.** *BMJ* 2003; 326:469-475.
3. Heikinheimo RJ, Inkovaara JA, Harju EJ et al. Annual injection of vitamin D and fractures of aged bones. *Calcif Tissue Int* 1992; 51(2):105-110.
4. Dawson-Hughes B, Dallal GE, Krall EA, Harris S, Sokoll LJ, Falconer G. Effect of vitamin D supplementation on wintertime and overall bone loss in healthy postmenopausal women [see comments]. *Annals of Internal Medicine* 1991; 115:505-512.
5. Chapuy MC, Arlot ME, Duboeuf F et al. Vitamin D3 and calcium to prevent hip fractures in the elderly women. *N Engl J Med* 1992; 327(23):1637-1642.
6. Dawson-Hughes B, Harris SS, Krall EA, Dallal GE. Effect of calcium and vitamin D supplementation on bone density in men and women 65 years of age or older [see comments]. *N Engl J Med* 1997; 337(10):670-676.
7. Chapuy MC, Pamphele R, Paris E et al. Combined calcium and vitamin D3 supplementation in elderly women: confirmation of reversal of secondary hyperparathyroidism and hip fracture risk: the Decalys II study. *Osteoporos Int* 2002; 13(3):257-264.
8. Pfeifer M, Begerow B, Minne HW, Abrams C, Nachtigall D, Hansen C. Effects of a short-term vitamin D and calcium supplementation on body sway and secondary hyperparathyroidism in elderly women. *J Bone Miner Res* 2000; 15(6):1113-1118.
9. Bischoff HA, Stahelin HB, Dick W et al. Effects of vitamin D and calcium supplementation on falls: a randomized controlled trial. *J Bone Miner Res* 2003; 18(2):343-351.
10. Janssen HC, Samson MM, Verhaar HJ. Vitamin D deficiency, muscle function, and falls in elderly people. *Am J Clin Nutr* 2002; 75(4):611-615.
11. Feskanich D, Willett WC, Colditz GA. Calcium, vitamin D, milk consumption, and hip fractures: a prospective study among postmenopausal women. *Am J Clin Nutr* 2003; 77(2):504-511.
12. Lau KW, Baylink DJ. Vitamin D Therapy of Osteoporosis: Plain Vitamin D Therapy Versus Active Vitamin D Analog (D-Hormone) Therapy. *Calcif Tissue Int* 1999; 65(4):295-306.
13. Bouillon RA, Auwerx JH, Lissens WD, Pelemans WK. Vitamin D status in the elderly: seasonal substrate deficiency causes 1,25-dihydroxycholecalciferol deficiency. *Am J Clin Nutr* 1987; 45(4):755-763.
14. Himmelstein S, Clemens TL, Rubin A, Lindsay R. Vitamin D supplementation in elderly nursing home residents increases 25(OH)D but not 1,25(OH)2D. *American Journal of Clinical Nutrition* 1990; 52:701-706.
15. Landin-Wilhelmsen K, Wilhelmsen L, Wilske J et al. Sunlight increases serum 25(OH) vitamin D concentration whereas 1,25(OH)2D3 is unaffected. Results from a general population study in Goteborg, Sweden (The WHO MONICA Project). *Eur J Clin Nutr* 1995; 49(6):400-407.
16. Vieth R, Ladak Y, Walfish PG. Age-Related Changes in the 25-Hydroxyvitamin D Versus Parathyroid Hormone Relationship Suggest a Different Reason Why Older Adults Require More Vitamin D. *J Clin Endocrinol Metab* 2003; 88(1):185-191.
17. Ishimura E, Nishizawa Y, Inaba M et al. Serum levels of 1,25-dihydroxyvitamin D, 24,25-dihydroxyvitamin D, and 25-hydroxyvitamin D in nondialyzed patients with chronic renal failure. *Kidney Int* 1999; 55(3):1019-1027.
18. Vitamin D3 at 90 or 7000 mcg weekly for 1 year: Responses of 25-hydroxyvitamin D, PTH, urine and plasma calcium. Rome, Italy: 2003.
19. Zehnder D, Bland R, Williams MC et al. Extrarenal Expression of 25-Hydroxyvitamin D(3)-1alpha-Hydroxylase. *J Clin Endocrinol Metab* 2001; 86(2):888-894.
20. Zineb R, Zhor B, Odile W, Marthe RR. Distinct, tissue-specific regulation of vitamin D receptor in the intestine, kidney, and skin by dietary calcium and vitamin D [In Process Citation]. *Endocrinology* 1998; 139(4):1844-1852.
21. Lefkowitz ES, Garland CF. Sunlight, vitamin D, and ovarian cancer mortality rates in US women. *Int J Epidemiol* 1994; 23(6):1133-1136.
22. Martinez ME, Giovannucci EL, Colditz GA et al. Calcium, vitamin D, and the occurrence of colorectal cancer among women. *J Natl Cancer Inst* 1996; 88(19):1375-1382.
23. Tangrea J, Helzlsouer K, Pietinen P et al. Serum levels of vitamin D metabolites and the subsequent risk of colon and rectal cancer in Finnish men. *Cancer Causes Control* 1997; 8(4):615-625.
24. Garland CF, Garland FC, Gorham ED. Can colon cancer incidence and death rates be reduced with calcium

- and vitamin D? *Am J Clin Nutr* 1991; 54(1 Suppl):193S-201S.
25. Emerson JC, Weiss NS. Colorectal cancer and solar radiation. *Cancer Causes & Control* 1992; 3:95-99.
26. Schwartz GG. Multiple sclerosis and prostate cancer: what do their similar geographies suggest? *Neuroepidemiology* 1992; 11:244-254.
27. Hanchette CL, Schwartz GG. Geographic patterns of prostate cancer mortality. Evidence for a protective effect of ultraviolet radiation. *Cancer* 1992; 70:2861-2869.
28. Ainsleigh HG. Beneficial effects of sun exposure on cancer mortality. [Review]. *Preventive Medicine* 1993; 22:132-140.
29. Grant WB. An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* 2002; 94(6):1867-1875.
30. Hayes CE. Vitamin D: a natural inhibitor of multiple sclerosis. *Proc Nutr Soc* 2000; 59(4):531-535.
31. McGrath J. Does 'imprinting' with low prenatal vitamin D contribute to the risk of various adult disorders? *Med Hypotheses* 2001; 56(3):367-371.
32. Hayes CE, Cantorna MT, DeLuca HF. Vitamin D and multiple sclerosis. *Proc Soc Exp Biol Med* 1997; 216(1):21-27.
33. McAlindon TE, Felson DT, Zhang Y et al. Relation of dietary intake and serum levels of vitamin D to progression of osteoarthritis of the knee among participants in the Framingham Study. *Ann Intern Med* 1996; 125(5):353-359.
34. Lane NE, Gore LR, Cummings SR et al. Serum vitamin D levels and incident changes of radiographic hip osteoarthritis: a longitudinal study. Study of Osteoporotic Fractures Research Group. *Arthritis Rheum* 1999; 42(5):854-860.
35. Rostand SG. Ultraviolet light may contribute to geographic and racial blood pressure differences. *Hypertension* 1997; 30(2 Pt 1):150-156.
36. Krause R, Buhring M, Hopfenmuller W, Holick MF, Sharma AM. Ultraviolet B and blood pressure [letter]. *Lancet* 1998; 352(9129):709-710. Ref Type: Journal (Full)
37. Pfeifer M, Begerow B, Minne HW, Nachtigall D, Hansen C. Effects of a short-term vitamin D(3) and calcium supplementation on blood pressure and parathyroid hormone levels in elderly women. *J Clin Endocrinol Metab* 2001; 86(4):1633-1637.
38. McMurray DN, Bartow RA, Mintzer CL, Hernandez-Frontera E. Micronutrient status and immune function in tuberculosis. *Annals of the New York Academy of Sciences* 1990; 587:59-69.
39. Muhe L, Lulseged S, Mason KE, Simoes EA. Case-control study of the role of nutritional rickets in the risk of developing pneumonia in Ethiopian children. *Lancet* 1997; 349(9068):1801-1804.
40. Hypponen E, Laara E, Reunanen A, Jarvelin MR, Virtanen SM. Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *Lancet* 2001; 358(9292):1500-1503.
41. Adachi JD, Bensen WG, Bianchi F et al. Vitamin D and calcium in the prevention of corticosteroid induced osteoporosis: a 3 year followup [see comments]. *J Rheumatol* 1996; 23(6):995-1000.
42. Adachi JD, Ioannidis G. Calcium and vitamin D therapy in corticosteroid-induced bone loss: what is the evidence? [In Process Citation]. *Calcif Tissue Int* 1999; 65(4):332-336.
43. Cooper L, Clifton-Bligh PB, Nery ML et al. Vitamin D supplementation and bone mineral density in early postmenopausal women. *Am J Clin Nutr* 2003; 77(5):1324-1329.
44. Vieth R. Vitamin D nutrition and its potential health benefits for bone, cancer, and other conditions. *Journal of Nutrition and Environmental Medicine* 2001; 11:275-291.
45. Barthel HR, Scharla SH. [Benefits beyond the bones -- vitamin D against falls, cancer, hypertension and autoimmune diseases]. *Dtsch Med Wochenschr* 2003; 128(9):440-446.
46. Holick MF. Vitamin D: A millenium perspective. *J Cell Biochem* 2003; 88(2):296-307.
47. Zittermann A. Vitamin D in preventive medicine: are we ignoring the evidence? *Br J Nutr* 2003; 89(5):552-572.
48. Vieth R, McCarten K, Norwich KH. Role of 25-hydroxyvitamin D3 dose in determining rat 1,25-dihydroxyvitamin D3 production. *American Journal of Physiology* 1990; 258(5 Pt 1):E780-9.
49. Vieth R, Milojevic S. Moderate vitamin D3 supplementation lowers serum 1,25-dihydroxy-vitamin D3 in rats. *Nutrition Research* 15[5], 725-731. 1995.
50. Vieth R, Bayley TA, Walfish PG, Rosen IB, Pollard A. Relevance of vitamin D metabolite concentrations in supporting the diagnosis of primary hyperparathyroidism. *Surgery* 1991; 110(6):1043-6; discussion 1046-7.
51. Bell NH. Endocrine complications of sarcoidosis. [Review]. *Endocrinology & Metabolism Clinics of North America* 1991; 20:645-654.
52. Morita R, Yamamoto I, Takada M, Ohnaka Y, Yuu I. [Hypervitaminosis D]. [Review] [Japanese]. *Nippon*

Rinsho - Japanese Journal of Clinical Medicine 1993; 51:984-988.

53. Dunn JF, Nisula BC, Rodbard D. Transport of steroid hormones: binding of 21 endogenous steroids to both testosterone-binding globulin and corticosteroid-binding globulin in human plasma. *J Clin Endocrinol Metab* 1981; 53(1):58-68.

54. Vieth R. Simple method for determining specific binding capacity of vitamin D-binding protein and its use to calculate the concentration of "free" 1,25-dihydroxyvitamin D. *Clin Chem* 1994; 40:435-441.

55. Haddad JG, Fraser DR, Lawson DE. Vitamin D plasma binding protein. Turnover and fate in the rabbit. *J Clin Invest* 1981; 67(5):1550-1560.

56. Vicchio D, Yergey A, O'Brien K, Allen L, Ray R, Holick M. Quantification and kinetics of 25-hydroxyvitamin D3 by isotope dilution liquid chromatography/thermospray mass spectrometry. *Biol Mass Spectrom* 1993; 22(1):53-58.

57. Nykjaer AWTE. The low-density lipoprotein receptor gene family: a cellular Swiss army knife? *TRENDS in Cell Biology* 12, 273-280. 2002.

Ref Type: Generic

58. Nykjaer A, Dragun D, Walther D et al. An endocytic pathway essential for renal uptake and activation of the steroid 25-(OH) vitamin D3. *Cell* 1999; 96(4):507-515.

59. Park EA. The therapy of rickets. *The J of the American Medical Association (JAMA)* 1940; 115(5):370-379.

60. Cooke R, Hollis B, Conner C, Watson D, Werkman S, Chesney R. Vitamin D and mineral metabolism in the very low birth weight infant receiving 400 IU of vitamin D. *Journal of Pediatrics* 1990; 116:423-428.

61. Pittard WB, Geddes KM, Hulseley TC, Hollis BW. How much vitamin D for neonates? *Am J Dis Child* 1991; 145(10):1147-1149.

62. Zeghoud F, Vervel C, Guillozo H, Walrant-Debray O, Boutignon H, Garabedian M. Subclinical vitamin D deficiency in neonates: definition and response to vitamin D supplements. *Am J Clin Nutr* 1997; 65(3):771-778.

63. Chesney RW. Vitamin D deficiency and rickets. *Rev Endocr Metab Disord* 2001; 2(2):145-151.

64. Dent CE, Smith R. Nutritional osteomalacia. *Q J Med* 1969; 38(150):195-209.

65. Blumberg RW, Forbes GB, Fraser D et al. The prophylactic requirement and the toxicity of vitamin D. *Pediatrics* 1963; 31:512-525.

66. Jablonski NG, Chaplin G. The evolution of human skin coloration. *J Hum Evol* 2000; 39(1):57-106.

67. Standing Committee on the Scientific Evaluation of Dietary Reference Intakes. *Dietary reference intakes: calcium, phosphorus, magnesium, vitamin D, and fluoride*. National Academy Press, 1997.

68. Vieth R, Fraser D. Vitamin D insufficiency: no recommended dietary allowance exists for this nutrient. *CMAJ* 2002; 166(12):1541-1542.

69. Vieth R. Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *Am J Clin Nutr* 1999; 69(5):842-856.

70. Marx SJ, Jones G, Weinstein RS, Chrousos GP, Renquist DM. Differences in mineral metabolism among nonhuman primates receiving diets with only vitamin D3 or only vitamin D2. *J Clin Endocrinol Metab* 1989; 69(1282):1282-1289.

71. Trang H, Cole DE, Rubin LA, Pierratos A, Siu S, Vieth R. Evidence that vitamin D3 increases serum 25-hydroxyvitamin D more efficiently than does vitamin D2. *The American Journal of Clinical Nutrition* 68, 854-848. 1998.

72. Bicknell F, Prescott F. Vitamin D. The antirachitic or calcifying vitamin. In: Bicknell F, Prescott F, editors. *Vitamins in Medicine*. London: Whitefriars Press, 1946: 630-707.

73. Norman AW. Problems relating to the definition of an international unit for Vitamin D and its metabolites. *J Nutrition* 1972; 102:1243-1246.

74. Horst RL, Napoli JL, Littledike ET. Discrimination in the metabolism of orally dosed ergocalciferol and cholecalciferol by the pig, rat and chick. *Biochem J* 1982; 204(1):185-189.

75. Marx SJ, Jones G, Weinstein RS, Chrousos GP, Renquist DM. Differences in mineral metabolism among nonhuman primates receiving diets with only vitamin D3 or only vitamin D2. *J Clin Endocrinol Metab* 1989; 69(6):1282-1290.

76. HEALTH & CONSUMER PROTECTION DIRECTORATE-GENERAL. **Opinion of the Scientific Committee on Food on the Tolerable Upper Intake Level of Vitamin D**. EUROPEAN COMMISSION, editor. SCF/CS/NUT/UPPLEV/38 Final, http://europa.eu.int/comm/food/fs/sc/scf/out157_en.pdf. 2002. Brussels, Belgium. 1-10-0030.

Ref Type: Report

77. Webb AR, DeCosta BR, Holick MF. Sunlight regulates the cutaneous production of vitamin D3 by causing its photodegradation. *J Clin Endocrinol Metab* 1989; 68:882-887.

78. Stamp TC. Factors in human vitamin D nutrition and in the production and cure of classical rickets. *Proc Nutr Soc* 1975; 34(2):119-130.

79. Davie MW, Lawson DE, Emberson C, Barnes JL, Roberts GE, Barnes ND. Vitamin D from skin: contribution to vitamin D status compared with oral vitamin D in normal and anticonvulsant-treated subjects. *Clin Sci* 1982; 63(5):461-472.
80. Chel VG, Ooms ME, Popp-Snijders C et al. Ultraviolet irradiation corrects vitamin D deficiency and suppresses secondary hyperparathyroidism in the elderly [In Process Citation]. *J Bone Miner Res* 1998; 13(8):1238-1242.
81. Holick MF. Environmental factors that influence the cutaneous production of vitamin D. *Am J Clin Nutr* 1995; 61(3 Suppl):638S-645S.
82. Haddock L, Corcino J, Vazquez Md. 25(OH)D serum levels in the normal Puerto Rican population and in subjects with tropical sprue and parathyroid disease. *Puerto Rico Health Sciences Journal* 1982; 1:85-91.
83. Haddad JG, Kyung JC. Competitive Protein-binding radioassay for 25-hydroxycholecalciferol. *Journal of Clinical Endocrinology* 1971; 33:992-995.
84. Better OS, Shabtai M, Kedar S, Melamud A, Berenheim J, Chaimovitz C. Increased incidence of nephrolithiasis in lifeguards in Israel. In: Massry SG, Ritz E, Jahreis G, editors. *Phosphate and Minerals in Health and Disease*. New York: Plenum Press, 1980: 467-472.
85. Matsuoka LY, Wortsman J, Hollis BW. Suntanning and cutaneous synthesis of vitamin D₃. *J Lab Clin Med* 1990; 116(1):87-90.
86. Mawer EB, Berry JL, Sommer-Tsilenis E, Beykirch W, Kuhlwein A, Rohde BT. Ultraviolet irradiation increases serum 1,25-dihydroxyvitamin D in vitamin-D-replete adults. *Miner Electrolyte Metab* 1984; 10(2):117-121.
87. Stamp TC, Haddad JG, Twigg CA. Comparison of oral 25-hydroxycholecalciferol, vitamin D, and ultraviolet light as determinants of circulating 25-hydroxyvitamin D. *Lancet* 1977; 1(8026):1341-1343.
88. Dent CE, Round JM, Rowe DJ, Stamp TC. Effect of chapattis and ultraviolet irradiation on nutritional rickets in an Indian immigrant. *Lancet* 1973; 1(7815):1282-1284.
89. Varghese M, Rodman JS, Williams JJ et al. The effect of ultraviolet B radiation treatments on calcium excretion and vitamin D metabolites in kidney stone formers. *Clin Nephrol* 1989; 31(5):225-231.
90. Falkenbach A, Unkelbach U, Boehm BO et al. Bone metabolism before and after irradiation with ultraviolet light. *Eur J Appl Physiol* 1993; 66(1):55-59.
91. Lawson DE, Sedrani SH, Douglas J. Interrelationships in rats of tissue pools of cholecalciferol and 25-hydroxycholecalciferol formed in u.v. light. *Biochem J* 1986; 233(2):535-540.
92. Lawson DE, Douglas J, Lean M, Sedrani S. Estimation of vitamin D₃ and 25-hydroxyvitamin D₃ in muscle and adipose tissue of rats and man. *Clin Chim Acta* 1986; 157(2):175-181.
93. Vieth R. Reply to FAJ Muskiet et al. *Am J Clin Nutr* 2001; 74(6):863-864.
94. Brouwer DA, van Beek J, Ferwerda H et al. Rat adipose tissue rapidly accumulates and slowly releases an orally-administered high vitamin D dose. *Br J Nutr* 1998; 79(6):527-532.
95. Clements MR, Davies M, Hayes ME et al. The role of 1,25-dihydroxyvitamin D in the mechanism of acquired vitamin D deficiency. *Clin Endocrinol (Oxf)* 1992; 37(1):17-27.
96. Preece MA, Tomlinson S, Ribot CA et al. Studies of vitamin D deficiency in man. *Q J Med* 1975; 44(176):575-589.
97. Dlugos DJ, Perrotta PL, Horn WG. Effects of the submarine environment on renal-stone risk factors and vitamin D metabolism. *Undersea Hyperb Med* 1995; 22(2):145-152.
98. Adams JS, Lee G. Gains in bone mineral density with resolution of vitamin D intoxication [see comments]. *Ann Intern Med* 1997; 127(3):203-206.
99. Koutkia P, Chen TC, Holick MF. Vitamin D intoxication associated with an over-the-counter supplement. *N Engl J Med* 2001; 345(1):66-67.
100. Vieth R, Pinto T, Reen BS, Wong MM. Vitamin D poisoning by table sugar. *Lancet* 2002; 359:672.
101. Vieth R, Chan A, Pollard A. 125I-RIA kit cannot distinguish vitamin D deficiency as well as a more specific assay for 25-hydroxyvitamin D. *Clinical Biochemistry* 1995; 28:175-179.
102. Stanbury SW, Mawer EB. The metabolism of a physiological dose of radioactive cholecalciferol (vitamin D₃) to its hydroxylated metabolites in man. *Clin Sci (Colch)* 1980; 58(6):523-535.
103. Mawer EB, Backhouse J, Holman CA, Lumb GA, Stanbury SW. The distribution and storage of vitamin D and its metabolites in human tissues. *Clin Sci* 1972; 43(3):413-431.
104. Rosenstreich SJ, Rich C, Volwiler W. Deposition in and release of vitamin D₃ from body fat: evidence for a storage site in the rat. *J Clin Invest* 1971; 50(3):679-687.
105. Montgomery JL, Parrish FC, Jr., Beitz DC, Horst RL, Huff-Lonergan EJ, Trenkle AH. The use of vitamin D₃ to improve beef tenderness. *J Anim Sci* 2000; 78(10):2615-2621.

106. Arunabh S, Pollack S, Yeh J, Aloia JF. Body fat content and 25-hydroxyvitamin d levels in healthy women. *J Clin Endocrinol Metab* 2003; 88(1):157-161.
107. Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF. Decreased bioavailability of vitamin D in obesity. *Am J Clin Nutr* 2000; 72(3):690-693.
108. Vieth R, Chan PC, MacFarlane GD. Efficacy and safety of vitamin D(3) intake exceeding the lowest observed adverse effect level. *Am J Clin Nutr* 2001; 73(2):288-294.
109. Barger-Lux MJ, Heaney RP, Dowell S, Chen TC, Holick MF. Vitamin D and its major metabolites: serum levels after graded oral dosing in healthy men. *Osteoporos Int* 1998; 8(3):222-230.
110. Pettifor JM, Bikle DD, Cavaleros M, Zachen D, Kamdar MC, Ross FP. Serum levels of free 1,25-dihydroxyvitamin D in vitamin D toxicity. *Ann Intern Med* 1995; 122(7):511-513.
111. Blank S, Scanlon KS, Sinks TH, Lett S, Falk H. An outbreak of hypervitaminosis D associated with the overfortification of milk from a home-delivery dairy. *Am J Public Health* 1995; 85(5):656-659.
112. Vieth R. The mechanisms of vitamin D toxicity. *Bone & Mineral* 1990; 11:267-272.
113. Narang NK, Gupta RC, Jain MK, Aaronson K. Role of vitamin D in pulmonary tuberculosis. *Journal of Association of Physicians of India* 1984; 32(2):185-186.
114. Munro I. Derivation of tolerable upper intake levels of nutrients. *Am J Clin Nutr* 2001; 74(6):865-867.
115. Yates AA. Process and development of dietary reference intakes: basis, need, and application of recommended dietary allowances. *Nutr Rev* 1998; 56(4 Pt 2):S5-S9.
116. Hathcock JN. Tolerable upper intake level of vitamin D. *American J Clinical Nutrition* 2001; in press.
117. Recommended Dietary Allowances. Seventh Revised Edition ed. Washington, D.C.: National Academy Press, 1968.
118. Heaney RP. Quantifying human calcium absorption using pharmacokinetic methods. *J Nutr* 2003; 133(4):1224-1226.
119. Jacobus CH, Holick MF, Shao Q et al. Hypervitaminosis D associated with drinking milk [see comments]. *New England Journal of Medicine* 1992; 326:1173-1177.
120. Bell NH. Renal and nonrenal 25-hydroxyvitamin D-1 α -hydroxylases and their clinical significance [In Process Citation]. *J Bone Miner Res* 1998; 13(3):350-353.
121. Barger-Lux MJ, Heaney RP. Effects of above average summer sun exposure on serum 25-hydroxyvitamin d and calcium absorption. *J Clin Endocrinol Metab* 2002; 87(11):4952-4956.
122. Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ. Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol. *Am J Clin Nutr* 2003; 77(1):204-210.
123. Muskiet FA, Dijck-Brouwer DJ, van d, V, Schaafsma A. Do we really need ≥ 100 μ g vitamin D/d, and is it safe for all of us? *Am J Clin Nutr* 2001; 74(6):862-863.
124. Lips P, Graafmans WC, Ooms ME, Bezemer PD, Bouter LM. Vitamin D supplementation and fracture incidence in elderly persons. A randomized, placebo-controlled clinical trial. *Ann Intern Med* 1996; 124(4):400-406.
125. Vieth R, Fraser D. Kinetic behavior of 25-hydroxyvitamin D-1-hydroxylase and -24- hydroxylase in rat kidney mitochondria. *J Biol Chem* 1979; 254(24):12455-12460.
126. Stene LC, Ulriksen J, Magnus P, Joner G. Use of cod liver oil during pregnancy associated with lower risk of Type I diabetes in the offspring. *Diabetologia* 2000; 43(9):1093-1098.
127. Eva JK. Vitamin D supplement in early childhood and risk for Type I (insulin-dependent) diabetes mellitus. The EURODIAB Substudy 2 Study Group. *Diabetologia* 1999; 42(1):51-54.
128. Fuller K. Lactose, rickets, and the coevolution of genes and culture. *Human Ecology* 2000; 28(3):471-477.
129. Eyles D, Brown J, Mackay-Sim A, McGrath J, Feron F. Vitamin d(3) and brain development. *Neuroscience* 2003; 118(3):641-653.
130. Mahon BD, Bemiss C, Cantorna MT. Altered cytokine profile in patients with multiple sclerosis following vitamin D supplementation. *FASEB J* 2001;837.4.
131. Embry AF, Snowdon LR, Vieth R. Vitamin D and seasonal fluctuations of gadolinium-enhancing magnetic resonance imaging lesions in multiple sclerosis. *Ann Neurol* 2000; 48(2):271-272.
132. Chan TY. Vitamin D Deficiency and Susceptibility to Tuberculosis. *Calcif Tissue Int* 2000; 66(6):476-478.
133. Douglas AS, Ali S, Bakhshi SS. Does vitamin D deficiency account for ethnic differences in tuberculosis seasonality in the UK? *Ethn Health* 1998; 3(4):247-253.
134. Lansdowne AT, Provost SC. Vitamin D3 enhances mood in healthy subjects during winter. *Psychopharmacology (Berl)* 1998; 135(4):319-323.

135. Gloth FM, III, Alam W, Hollis B. Vitamin D vs broad spectrum phototherapy in the treatment of seasonal affective disorder. *J Nutr Health Aging* 1999; 3(1):5-7.
136. Harris S, Dawson-Hughes B. Seasonal mood changes in 250 normal women. *Psychiatry Res* 1993; 49(1):77-87.
137. Al Allaf AW, Mole PA, Paterson CR, Pullar T. Bone health in patients with fibromyalgia. *Rheumatology (Oxford)* 2003; .
138. Garland CF, Garland FC, Gorham ED. Calcium and vitamin D. Their potential roles in colon and breast cancer prevention. *Ann N Y Acad Sci* 1999; 889:107-19:107-119.
139. Grant WB. An estimate of excess cancer mortality in the US due to inadequate exposure to solar UV-B radiation (abstract). *Photodermatol Photoimmunol Photomed* 2001; 17:142.
140. Schwartz GG, Wang MH, Zang M, Singh RK, Siegal GP. 1 alpha,25-Dihydroxyvitamin D (calcitriol) inhibits the invasiveness of human prostate cancer cells. *Cancer Epidemiol Biomarkers Prev* 1997; 6(9):727-732.
141. Schwartz GG, Whitlatch LW, Chen TC, Lokeshwar BL, Holick MF. Human prostate cells synthesize 1,25-dihydroxyvitamin D3 from 25-hydroxyvitamin D3 [In Process Citation]. *Cancer Epidemiol Biomarkers Prev* 1998; 7(5):391-395.
142. Hsu JY, Feldman D, McNeal JE, Peehl DM. Reduced 1alpha-hydroxylase activity in human prostate cancer cells correlates with decreased susceptibility to 25-hydroxyvitamin D3-induced growth inhibition. *Cancer Res* 2001; 61(7):2852-2856.
143. Vieth R, Dogan M, Cole DEC et al. Vitamin D3 at 90 or 700 mcg weekly for 1 year: responses of 25(OH)D, PTH, urine and plasma calcium. *European Calcified Tissue Society, Rome, May, 2003 Abstract. 2003. : Abstract*
144. Harris S. Can vitamin D supplementation in infancy prevent type 1 diabetes? *Nutr Rev* 2002; 60(4):118-121.
145. Heaney RP, Davies KM, Chen TC, Holick MF, Barger-Lux MJ. Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol. *Am J Clin Nutr* 2003; 77(1):204-210.
146. Marx SJ, Jones G, Weinstein RS, Chrousos GP, Renquist DM. Differences in mineral metabolism among nonhuman primates receiving diets with only vitamin D3 or only vitamin D2. *J Clin Endocrinol Metab* 1989; 69(6):1282-1290.
147. Jones G, Byrnes B, Palma F, Segev D, Mazur Y. Displacement potency of vitamin D2 analogs in competitive protein-binding assays for 25-hydroxyvitamin D3, 24,25-dihydroxyvitamin D3, and 1,25-dihydroxyvitamin D3. *J Clin Endocrinol Metab* 1980; 50(4):773-775.
148. Mawer EB, Jones G, Davies M et al. Unique 24-hydroxylated metabolites represent a significant pathway of metabolism of vitamin D2 in humans: 24-hydroxyvitamin D2 and 1,24-dihydroxyvitamin D2 detectable in human serum. *J Clin Endocrinol Metab* 1998; 83(6):2156-2166.
149. Holmberg I, Berlin T, Ewerth S, Bjorkhem I. 25-Hydroxylase activity in subcellular fractions from human liver. Evidence for different rates of mitochondrial hydroxylation of vitamin D2 and D3. *Scand J Clin Lab Invest* 1986; 46(8):785-790.
150. Guo YD, Strugnell S, Back DW, Jones G. Transfected human liver cytochrome P-450 hydroxylates vitamin D analogs at different side-chain positions. *Proceedings of the National Academy of Sciences of the United States of America* 1993; 90:8668-8672.
151. Harris SS, Dawson-Hughes B, Perrone GA. Plasma 25-hydroxyvitamin D responses of younger and older men to three weeks of supplementation with 1800 IU/day of vitamin D. *J Am Coll Nutr* 1999; 18(5):470-474.